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Critical Review

Plastic particle ingestion by wild freshwater fish: a critical review

France Collard, Johnny Gasperi, Geir Wing Gabrielsen, and Bruno Tassin

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1 Plastic particle ingestion by wild freshwater fish: a critical review

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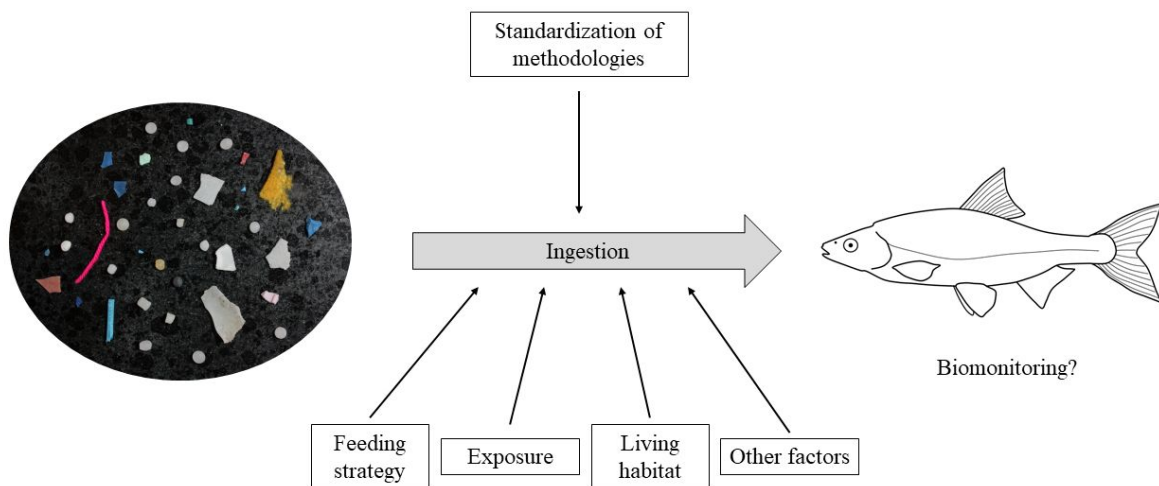
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25 Graphical abstract

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29 Abstract

30

31 Plastic pollution, especially microplastics (MP) pollution, is a hot topic in both mainstream
32 media and scientific literature. Although rivers are potentially the major transport pathway of
33 this pollution to the sea, plastic contamination in freshwater bodies is comparatively
34 understudied. Microplastic pollution in freshwater fish is of growing interest, and while few
35 studies exist, discrepancies do occur in the sampling, extraction, and identification of MP and
36 in the expression of the results. Even though those differences hamper comparisons between
37 some studies, a comparative work has been performed to identify the factors influencing MP
38 ingestion by fish and consequently to target potential ecological traits that can be used to
39 monitor species. Monitoring plastic ingested by fish will give relevant ecological information
40 on MP pollution. This review focuses on MP ingestion by wild freshwater and estuarine fish.
41 In addition to providing an overview of the existing data concerning contamination levels in
42 wild freshwater fish, we aimed to (1) propose several overall recommendations on the
43 methodologies applicable to all biota, (2) compare MP contamination levels in fish and in their
44 environment, and (3) determine which parameters could help to define fish species for
45 monitoring.

46

47

48 1. Introduction

49

50 Due to their exponentially increasing production since the 1950s,¹ plastic materials are polluting
51 all types of environments: marine surface waters,^{2,3} deep-sea sediments,^{4,5} arctic sea ice,^{6,7}
52 soils,⁸ and even air.^{9,10} As an expected and direct consequence, the number of species or taxa
53 exposed to plastic pollution is alarming.¹¹ Plastic pieces of all sizes are found in the

54 environment, and thus are ingested by many different organisms: marine mammals,¹²⁻¹⁴ marine
55 and terrestrial birds,¹⁵⁻¹⁷ crustaceans,^{18,19} worms,²⁰ and fish.²¹⁻²³ Scientific research is currently
56 more focused on the marine environment. The reasons for this focus are probably due the high
57 economic value of marine resources and abundant funding allocated to marine research and
58 monitoring. In particular, microplastics (MP), which are plastic pieces with sizes less than 5
59 mm,²⁴ are well known to pollute seas and oceans. Numerous laboratory studies have shown that
60 this exposure leads to plastic ingestion by various organisms and is associated with some
61 negative impacts,²⁵ such as neurotoxicity,²⁶ a change in swimming behavior²⁷ and reduction of
62 predatory performances.²⁸

63 Human populations are closely linked to water from both marine and freshwater sources. A
64 considerable fraction of the human population lives in the near-coastal zone.²⁹ Approximately
65 half of the world's population lives within 3 km of a freshwater body and only 10% of the
66 population lives farther than 10 km away.³⁰ However, given that proximity, between 1.15 and
67 2.41 million tons of plastics are released into the oceans via rivers each year.³¹ Conventional
68 wastewater treatment plants may act as an MP source to rivers,^{32,33} and inappropriate waste
69 management³⁴ combined with a high population density³⁵ may both be positively correlated
70 with riverine plastic loads.

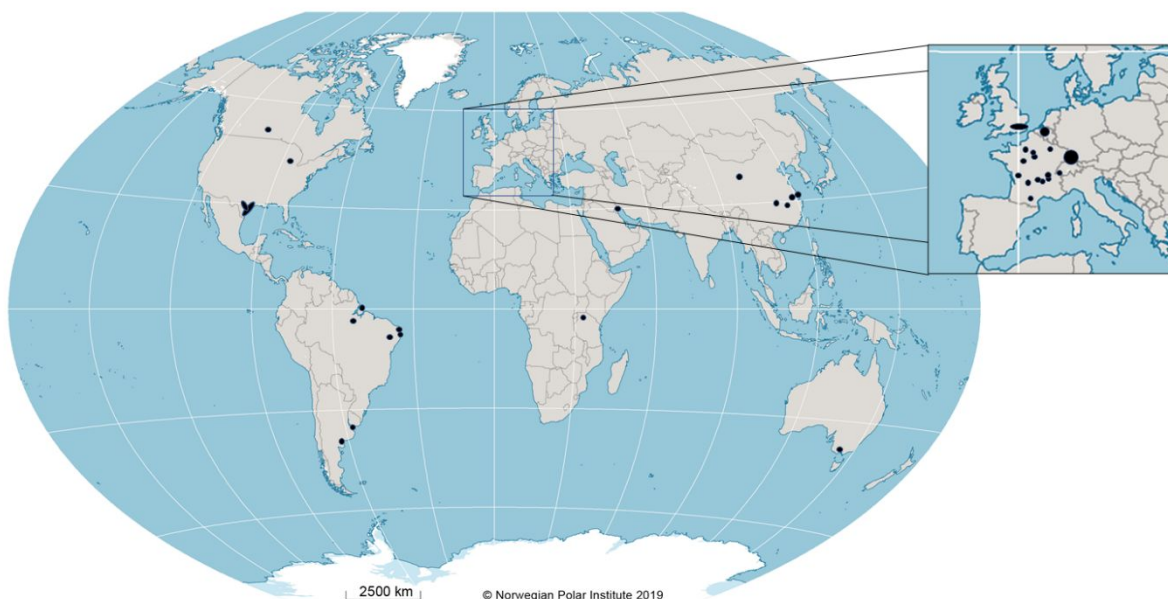
71 Increasing number of studies have revealed that freshwaters are contaminated, sometimes
72 largely by plastic, regardless of its size.³⁶⁻³⁹ The number of studies about MP pollution and its
73 impacts is growing (in March 2019, searching the Web of Science for “plastic marine pollution”
74 returned 3 results in 2000 and 351 results in 2018), but a small number of these studies deal
75 with a freshwater environment (0 out of the 3 results found in 2000 and 42 out of the 351 results
76 found in 2018). The same observation can be made for fish; searching the Web of Science for
77 the keywords “fish plastic pollution” gave 4 results in 2000 and 99 in 2018, in which 0 and 10
78 results concern freshwaters, respectively. Despite the large amounts of plastic debris input into

79 seas and oceans by rivers, the interactions between these debris and the biota of these
80 ecosystems are poorly studied. Nevertheless, some discrepancies in performed protocols do
81 occur. Among others, the sampling size is frequently too small, and MP identification often
82 relies on visual sorting. In addition, the results are reported in one or two units while at least
83 three are used in publications. Those discrepancies lead to difficulties in comparing data.

84 Fish are the main taxa studied with regard to MP ingestion in freshwater environments⁴⁰ and
85 constitute an economically, ecologically and highly diversified group.^{41,42} Microplastic
86 ingestion by freshwater fish has been studied worldwide, but these studies have although
87 focused on limited regions (Fig. 1) in comparison with the studies of plastic ingested by wild
88 marine fish.⁴³ In total, the studies focusing on freshwater fish, while few, have described MP
89 ingestion by more than 200 species. This large number indicates that some studies have
90 collected few samples. Few published papers have studied the interactions between MP and
91 freshwater biota compared to the number of marine studies, although scientific interest in MP
92 ingestion by freshwater fish is rising. We thus believe that a summary of recommendations for
93 the methods and expression of results and factors leading to MP ingestion by freshwater fish
94 would be useful for the scientific community, especially regarding monitoring. Three main
95 factors have been investigated: the MP levels in abiotic compartments, the living habitat and
96 the feeding strategy. Other factors could have been reviewed, but there are too few studies to
97 allow for a reliable discussion. Exposure through abiotic compartments could be directly
98 correlated with the ingestion of MP. As further discussed in this review, MP ingestion does not
99 always depend on abiotic compartment exposure. Additionally, there is not a clear relationship
100 between MP ingestion and the living habitat and feeding strategy. Some studies have found that
101 benthic or demersal fish ingested more MP,^{44,45} while others reported the contrary.²²

102 Monitoring and assessment are essential steps towards addressing specific questions about
103 marine litter, including MP. Monitoring and assessment are needed to assess the state or level

104 of pollution and provide objective information to design mitigation measures as well as assess
105 their effectiveness and promote adaptive management.⁴⁶ Recommendations for defining
106 indicator fish species are very few but crucial to improve the comparability between studies.
107



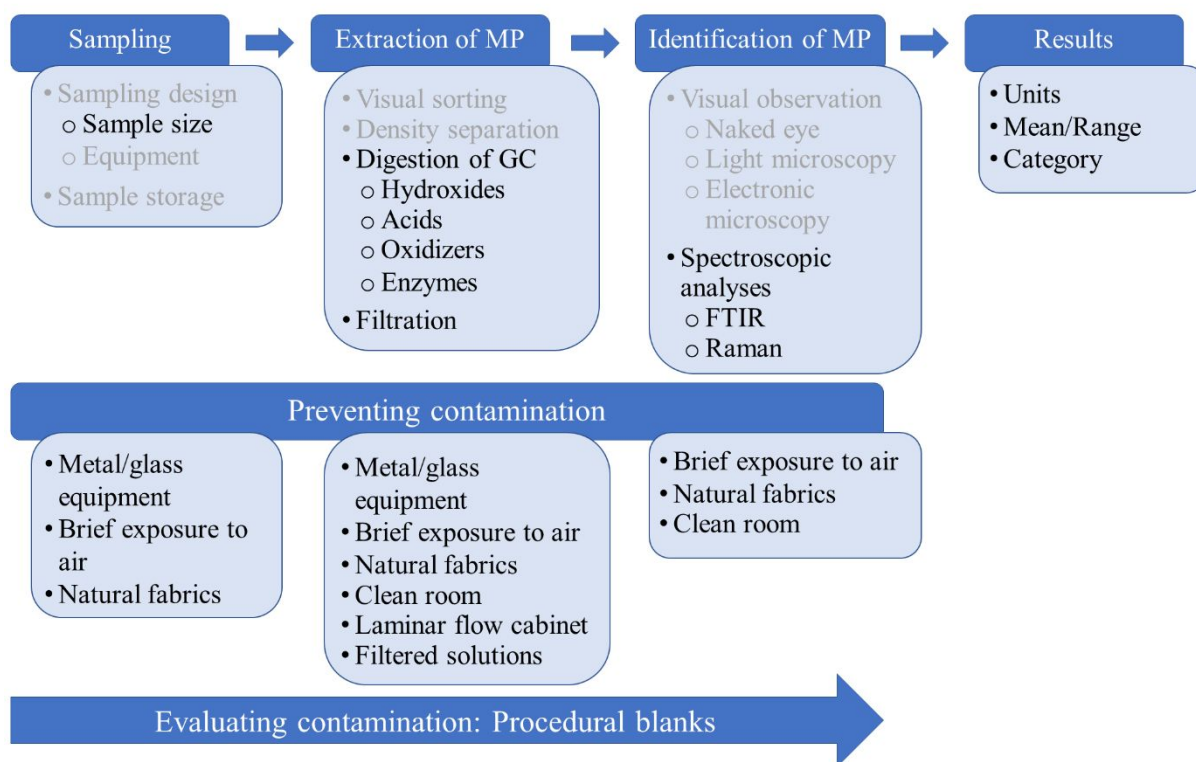
108
109 Figure 1. Global overview of the studies performed on MP ingestion by wild freshwater fish.
110
111 In this critical review, we aim to (1) propose several recommendations for the harmonization
112 of methodologies, which is applicable not only to MP contamination in freshwater fish studies
113 but also to all biota, (2) compare MP contamination levels in fish and their environment, and
114 subsequently (3) determine which parameters could help to define fish species for monitoring.
115 Our work is based on the occurrence of MP in the guts of fish. The translocation of plastic
116 particles to other organs is beyond the scope of this review but should be considered in further
117 studies.
118

119 2. Methodologies for the study of MP contamination in fish guts

120

121 Globally, studies on MP ingestion by fish, both marine and freshwater, follow a general pattern
122 regarding their methodology, which include sampling, gut content (GC) extraction, MP
123 extraction (visual sorting or chemical treatment, which is not always performed), and
124 identification (including spectroscopic analyses which are not always performed). No specific
125 method has been defined as the method to be used in fish GC analyses. In addition, this general
126 pattern is not specific to studies about fish, as it is also used for other organisms or even for
127 water or sediment matrices. In the two latter cases, density separation is often chosen to carry
128 out the MP extraction. Whatever the matrix, discrepancies in methodologies occur, including
129 in the study of MP in the GCs of wild freshwater fish. There are variations in all the steps used
130 to extract and identify MP. In the next sections, we highlight the main issues of the key protocol
131 steps used to extract MP from GCs (Fig. 2) and propose some recommendations to increase the
132 comparability of studies for future research. We will discuss the following main steps: the
133 sampling, extraction, and identification of MP and the expression of the results. Within those
134 steps, we highlight some parameters that we think are of great importance including the
135 sampling size, method for digesting the GCs, definition of the size threshold for the extracted
136 particles, spectroscopic analyses, and the way results are provided. In addition, some
137 recommendations for preventing and evaluating the contamination will be proposed. We
138 selected the most important criteria, as other reviews have already focused on methodologies
139 for the sampling, extraction and analyses of MP in biota or abiotic compartments.^{40,47,48}

140



141

142 Figure 2. Flowchart of the main steps used to analyze microplastics in fish gut contents. The
 143 steps shown in black are discussed in this review. GCs: gut contents, FTIR: Fourier transform
 144 infrared.

145

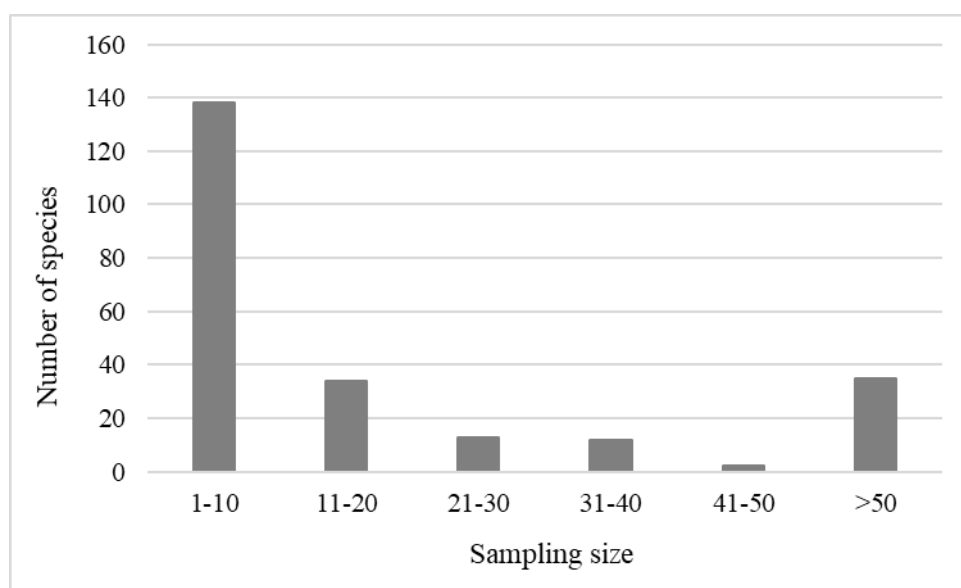
146 2.1. Sampling: Sample size

147

148 Among all the species studied, more than twenty have been analyzed at least twice, and most
 149 of these species have been studied thanks to studies conducted in South America.⁴⁹⁻⁵² In
 150 addition, when considering a sampling of at least 10 individuals per species, only seven out of
 151 twenty species can be spatially compared. A review focusing on wild marine fish found that the
 152 detection of plastic ingestion was positively correlated with an increased sample size (up to
 153 $n=10$).⁴³ In many studies reviewed here, the number of individuals representing each species
 154 was low ($n<10$), which is obviously too small to define solid trends in MP contamination. In
 155 several publications, data were given for one or two individuals, limiting the representativeness.

156 Given that a single dataset is available and the number of fish sampled is usually low for most
157 studied species, very local one-time events may influence the results and impair the global
158 understanding of MP ingestion by fish. A large sample size is highly important since it will
159 provide enough data to perform reliable statistical analyses. Previously, a threshold of 50
160 individuals has been defined as sufficiently reliable regarding to achieve statistical power,^{48,53,54}
161 while 40 specimens were considered to be an adequate sample size for the monitoring of plastic
162 ingestion by northern fulmars.⁵⁵ Fig. 3 shows that most studies did not sample more than 50
163 specimens per species. This chart highlights the need for a better sampling design to statistically
164 support the results.

165



166

167 Figure 3. Number of species for each category of sampling size. This chart is based on studies
168 shown in Table 1.

169

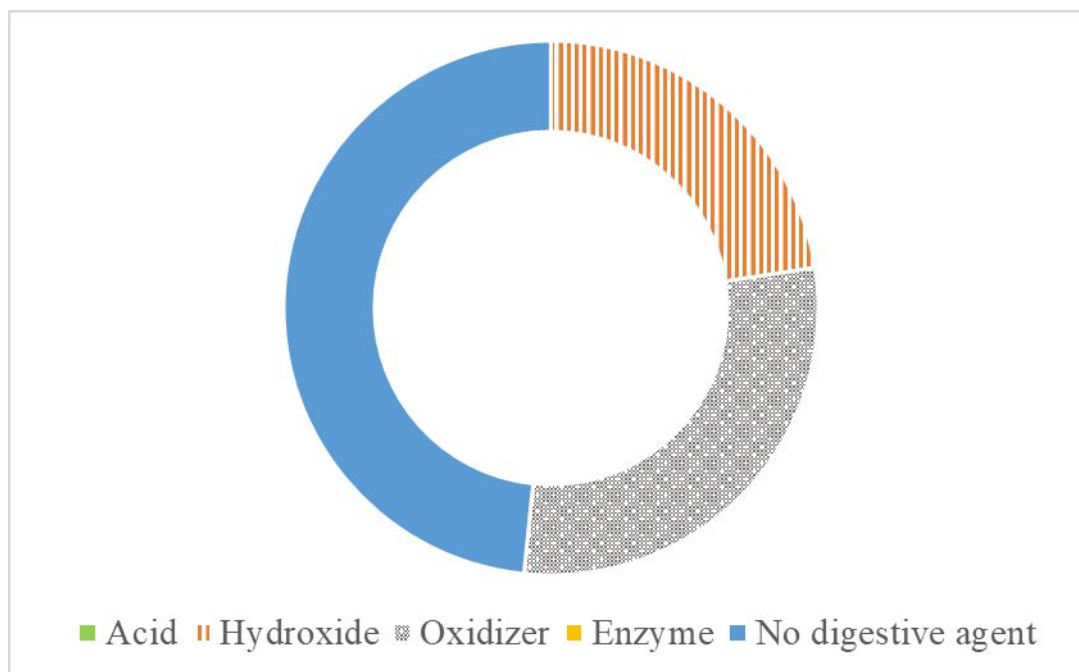
170 2.2. Extraction of MP: Digestion of fish gut contents

171

172 The digestion of organic matter is not mandatory but recommended,⁴⁶ especially when aiming
173 to isolate MP from biota. Several methods exist regarding GCs, which are indiscriminate for

174 marine and freshwater fish species. Fig. 4 shows that when present, the most common agents
175 are oxidizers (*e.g.*, H₂O₂ and NaClO)^{44,56,57} and hydroxides (*e.g.*, KOH).^{58,59} No studies reported
176 acids as the main digesting agent, which is most likely because acids alter some polymers, such
177 as polyethylene terephthalate (PET), high density polyethylene (HDPE) and polyamide (PA).
178 Acids can also lead to suboptimal digestion.^{58,60-62} Remarkably, to our knowledge, no studies
179 have used enzymes as digesting agents yet. While the efficiency of such agents has been
180 assessed,⁶²⁻⁶⁴ the cost and duration of this treatment might discourage researchers. Recently,
181 von Friesen et al.⁶⁵ proposed a protocol more efficient than that using KOH with the convenient
182 advantages of being less time-consuming and commercially available at a low price. This
183 protocol has been thoroughly tested using bivalves but suits a broader range of organisms
184 according to the authors. No studies on fish GCs have used Fenton's agent either, which is
185 probably due to the efficiency of the previously cited methods. This agent does not affect plastic
186 polymers and reduces the sample preparation time.⁶⁶ Both enzymes and Fenton's agent could
187 be a solution to achieving the efficient, rapid and inexpensive digestion of the organic matter in
188 the GCs of freshwater fish but could probably also be used in other biological matrices such as
189 the liver. Some GCs can be very fatty and challenging to digest. No study has identified a means
190 of treating those samples properly. Finding an easy way to process fatty GCs would be useful
191 for further research on fish gut contamination.

192



193

194 Figure 4. Proportion of studies according to the main digestive agent used.

195

196 Only a couple of studies have reported data on all types of anthropogenic particles, *i.e.*, small
197 pieces created or handled by humans, such as plastics, dyed particles or textile fibers.^{57,67}

198 Indeed, studies have only focused on plastic materials and have developed protocols for that
199 purpose. To our knowledge, few studies have tested the protocols used to extract MP from biota

200 on natural or modified natural materials such as cellulose or cellulose acetate.^{58,68,69} Dehaut et
201 al.⁵⁸ tested three extraction protocols on cellulose acetate materials, in which two contained

202 hydroxides (NaOH & KOH) and one contained a combination of a hydroxide and an oxidizer
203 (NaOH & K₂S₂O₈). All the protocols led to a mass reduction and a size and shape modification

204 of the cellulose acetate while two of them allowed for identification by Raman spectroscopy.

205 On the other hand, cellulose fibers, along with plastic polymers, were tested with an oxidizing
206 agent and a combination of an oxidizer and an acid (NaClO & HNO₃).⁶⁸ Cellulose fibers were

207 left unaffected by both treatments and Raman spectroscopy could successfully be performed.

208 A potassium hydroxide-based protocol has also been tested on several natural materials.⁶⁹

209 Cellulose acetate has been found to lose a significant part of its mass after treatment, which is

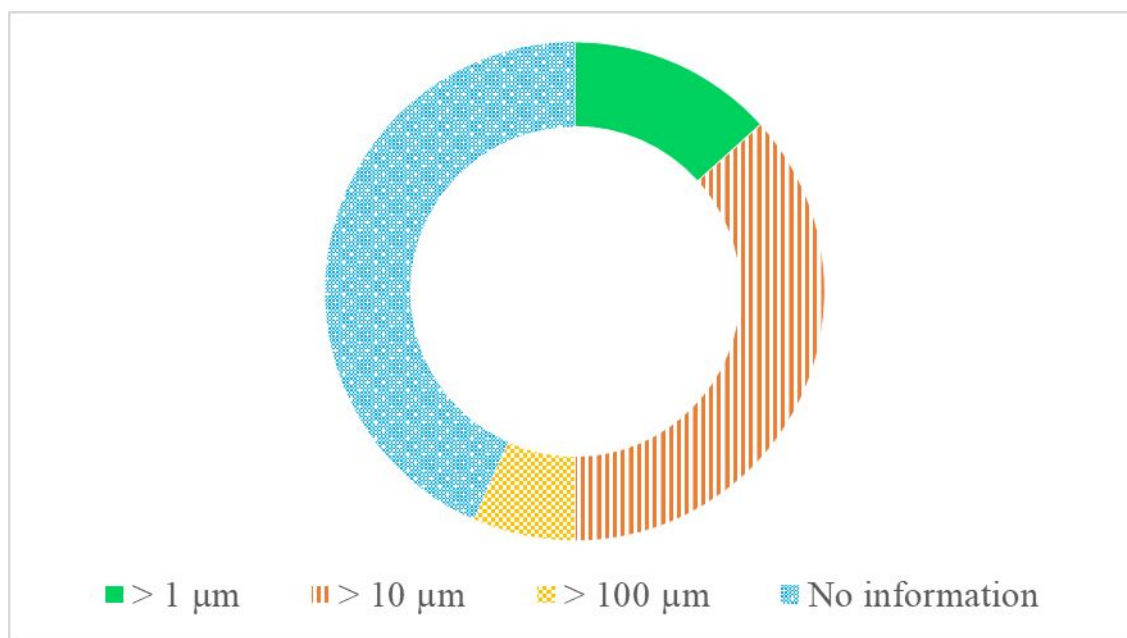
210 similar to the study by Dehaut et al.⁵⁸ Some natural materials, such as sheep wool, were
211 dissolved, while others showed a resistance to KOH. The extraction method should then be
212 carefully chosen when textile fibers are targeted.

213 Data on anthropogenic particles made from natural materials such as wool and cotton should
214 be included in further studies. Laboratory studies have mainly focused on plastic materials.
215 Textile fibers, independent of their composition, are found in high quantities in the
216 environment^{3,70} but their impacts are unknown. These particles contain additives and dyes^{16,70,71}
217 and may constitute a threat to the environment^{72,73}, aquatic organisms and human health.^{71,74}
218 All anthropogenic particles, regardless of their size, should therefore be studied in both in situ
219 and in vitro studies.

220

221 2.3. Extraction of MP: Filtration and targeted MP sizes

222



223

224 Figure 5. Proportion of studies according to their lowest limit of MP extracted. Based on Table
225 1.

226

227 The minimum size of the targeted particles is not always mentioned (Fig. 5). Generally, the
228 particle isolation process includes a filtration/sieving step, in which a precise mesh size
229 practically defines the threshold for particles to be further analyzed. In several studies, that
230 mesh size is close to the micrometer scale (Fig. 5). Although this mesh size perfectly fits the
231 microplastic size definition, from 5 mm down to a few microns,⁷⁵ the limiting factor is then the
232 spectroscopic analyses. Depending on the technique and equipment used, the size threshold
233 allowing for the analysis of a particle varies. For instance, when dealing with particles between
234 1 and 20 μm in length, Raman spectroscopy is recommended.⁷⁶ However, if the aim of the study
235 is to investigate direct ingestion by fish, it might not be worth studying such small MP. Adult
236 fish are unlikely to ingest micron-sized particles because these particles will probably be ejected
237 into the surrounding water from the branchial system, which is not adapted to retain such small
238 particles.⁷⁷ The filtration step could then be made easier by using larger mesh sizes depending
239 on the studied species, mainly if no digestion step is performed. However, if the transfer of MP
240 from prey is considered in the study, looking at smaller particles might be of interest. Regarding
241 filtering fish, their filtration system defines the size limit of particles that are retained and further
242 swallowed. For those species, correlating the filtration-limited size to the size of MP ingested
243 might be a way to discriminate MP coming from prey (indirect ingestion) and MP ingested
244 through feeding (direct ingestion). This assumption has to be assessed by future studies. A size
245 of 5 μm could be the lowest limit of investigated MP sizes, as it almost perfectly covers the MP
246 size definition, which comprises direct and indirect MP ingestion and allows for reliable
247 spectroscopic analyses. We encourage authors to provide at least the size classes of the ingested
248 particles, as well as the size distribution. This information will allow for the comparison of
249 those size classes according to the minimal size targeted and will provide baseline data for
250 studies further dealing with the uptake of ingested MP by other organs and tissues.

251

2.4. Identification of MP: Spectroscopic analyses

Half of the studies on plastics in freshwater fish used spectroscopic analyses to confirm the plastic composition of the particles found, and most of these studies are the oldest (Table 1). When considering other matrices such as sediment or water, such techniques are often set aside.⁷⁶ Thus, these techniques only rely on visual observation, which may bias the results.⁷⁸⁻⁸⁰ Visual observations with a stereomicroscope tend to underestimate the amount of microplastic fragments and, conversely, tend to overestimate the amount of MP fibers compared to the results obtained by Fourier transform infrared (FTIR) analyses.⁷⁹ Markic et al.⁴³ found that plastic was detected more often when a chemical digestion-coupled spectroscopic analyses was performed compared to visual methods alone. The smaller the particles are, the more difficult it is to distinguish them from biological tissues and to assess their synthetic composition with the naked eye. The ‘hot point-test’ is also sometimes used to determine the plastic composition of particles.^{18,81} This test consists of touching a particle with a hot needle and observing whether the needle leaves a mark on the particle.^{48,82} Although convenient and cheap, this technique should not be solely used for identification but used as a complement to visual sorting prior to spectroscopic analyses.⁴⁸ In a report, the GESAMP group of experts⁴⁰ stated that spectroscopy is recommended for particles smaller than 1 mm. Consequently, using either Raman or FTIR spectroscopy when working with microplastic pollution is highly recommended.^{46,80} Both techniques usually allow for precisely identify the particle composition to be precisely identified and have the strong advantage of being nondestructive in contrast with other techniques such as pyrolysis-gas chromatography coupled with mass spectrometry (py-GC/MS) or thermogravimetric analysis coupled with solid-phase extraction (TGA-SPE).⁸³⁻⁸⁵ These techniques are complementary, as they give different information but using two or three of these techniques is obviously expensive and time-consuming. Therefore, depending on the

277 information needed or the type of particle, researchers have to choose the best technique. Raman
278 spectroscopy constitutes a better choice if results about additives are needed. Indeed Raman
279 spectroscopy can provide additional information on dyes, for example, and will identify the
280 composition of small particles ($< 20 \mu\text{m}$) that FTIR will miss.^{79,84,85} On the other hand, FTIR
281 spectroscopy is less time-consuming and may allow for the identification of black particles that
282 lead to fluorescence issues in Raman spectroscopy.⁸⁴ All these techniques are expensive and
283 time consuming; thus, a random subsample is often collected prior to the analyses, and the
284 results are extrapolated to the total number particles found.^{10,44,86-88} This random subsample is
285 usually poorly described and might result from subjective choices, especially regarding the
286 sizes and colors. As selecting the particles may lead to bias, we instead recommend selecting a
287 zone covering both the center and the edge of the filter (a half or a quarter) used during MP
288 extraction.

289 Spectroscopic analyses provide useful and reliable data on particles. However, part of this data
290 is often set aside because studies focus only on plastic polymers (Table 1). As explained in a
291 previous section, we encourage researchers to provide all information, even on natural and
292 semisynthetic materials.

293

294 2.5. Expression of the results

295

296 We recorded three different units of MP contamination in GCs, *i.e.*, the percentage of
297 contaminated individuals, the number of MP per individual and the number of MP in one gram
298 of the GCs. Additionally, units of contamination differ between studies making comparisons
299 impossible. All three units are easy to calculate and, although the most common (Table 2), the
300 percentage of occurrence is not the most representative unit. This unit gives an overview of the
301 number of fish having ingested at least one piece of plastic in a given time but does not give

302 any valuable quantitative information. The number of MP per gram of GCs could be considered
303 to be the most accurate unit but greatly depends on the fullness of the stomach or gut. As it is
304 not clear whether MP have a greater retention time than food in the digestive tract,^{89,90}
305 expressing the quantity of MP per mass of gut contents is not the most consistent option. It is
306 instead preferable to express the results as the number of MP per individual and for comparison
307 purposes, we recommend authors and researchers to report the results with the three units
308 described in this review.

309

310 2.6. Procedural blanks

311

312 Procedural blanks (PBs), also termed controls or negative controls,^{47,91} are used in all procedure
313 steps with the reagents only, and the sample in itself is excluded. Few studies that we have
314 reviewed made PBs although most of the recent studies did. Even if contamination is prevented
315 as best as possible, including PBs is recommended to evaluate possible contamination of the
316 whole treatment protocol^{46,48} and, if needed, to adapt the results according to those blank
317 data.^{18,82,91} Plastic contamination levels depend on many parameters, including the type of
318 equipment used for sampling, the extraction and the analyses.⁹² Given that microplastic fibers
319 are also found in the air,^{9,10} tap water⁹³ and table salt,^{94,95} in which the latter is used in
320 laboratories to perform density separation, PBs are necessary to evaluate the contamination
321 levels in environmental samples. Moreover, we recommend including some PBs before starting
322 the sample treatment. This will give an overview about the contamination level in the working
323 environment, which can be then compared to the expected levels in the samples. This step could
324 be critical if minimally contaminated samples are to be processed. In this case, PB levels would
325 easily exceed what may be found in the samples, leading to an impossible interpretation of the
326 results. Depending on the studies and equipment used, PB contamination can be important: 50

327 times higher than plastics within the samples,⁹⁶ or low: less than 5% of all plastics found in the
328 samples.⁴⁴ However, if PB levels are assessed first, precautions and decisions can be made to
329 prevent the samples from being highly contaminated during the treatment.

330

331 Table 1. Summary of studies reporting MP ingestion by wild freshwater fish and their methodologies.

No. sp.	n/sp.	Country	Water body	Targeted particles		Extracted particle size (µm)	Visual sorting	Digesting agent				Spectroscopic analyses		Procedural blanks	Reference
				AP	PL			Hyd.	Acid	Enz.	Ox.	Rn	FTIR		
3	60, 60, 62	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Possatto et al. ⁹⁷
2	239 & 330	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Dantas et al. ⁴⁹
3	240, 141, 44	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Ramos et al. ⁵⁰
1	186	France	R	-	X	> 1.2	X	-	-	-	-	-	-	-	Sanchez et al. ⁹⁸
2	20	Tanzania	L	-	X	> 500	X	X	-	-	-	-	X	-	Biginagwa et al. ⁹⁹
4	10	Switzerland	L	-	X	?	X	-	-	-	-	-	-	-	Faure et al. ¹⁰⁰
44	1-67	U.S.A.	R & E	-	X	?	X	-	-	-	-	-	X	-	Phillips & Bonner ¹⁰¹
1	530	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Ferreira et al. ¹⁰²
2	318 & 118	U.S.A.	R	-	X	> 53	X	-	-	-	-	-	-	-	Peters & Bratton ¹⁰³
5	10-75	Canada	R	-	X	> 5	X*	-	-	-	X	-	-	X	Campbell et al. ⁹⁶
6	20-40	China	L	-	X	> 5	X	-	-	-	X	-	X	X	Jabeen et al. ⁴⁴
2	10 & 66	England	R & E	-	X	?	X	-	-	-	-	-	X	-	McGoran et al. ¹⁰⁴
11	1-21	Argentina	E	-	X	?	X*	-	-	-	X	-	-	-	Pazos et al. ⁵⁶
1	48	Brazil	R	-	X	> 63	X	-	-	-	-	-	-	-	Silva-Cavalcanti et al. ¹⁰⁵
27	2-215	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Vendel et al. ⁵¹
13	1-6	China	R	-	X	> 1.2	X*	X	-	-	-	X	-	-	Zhang et al. ⁵⁹
1	60	France	R	X	X	> 5	-	-	-	-	X	X	-	X	Collard et al. ⁵⁷
1	64	England	R	X	X	> 1.2	X	-	-	-	-	X	-	-	Horton et al. ⁶⁷
46	1-16	Brazil	E	-	X	?	X	-	-	-	-	-	X	-	Pegado et al. ⁵²
4	4-17	Iran	E	-	X	?	X	X	X	-	X	-	-	X ⁺	Abbasi et al. ¹⁰⁶
11	1-17	U.S.A.	R	-	X	> 0.45	X*	-	-	-	X	-	X	X	McNeish et al. ⁹¹
1	10	China	L	-	X	> 200	X	X	-	-	-	X	-	X	Xiong et al. ¹⁰⁷
16	1-63	Brazil	R	-	X	?	X	-	-	-	-	-	X	-	Andrade et al. ¹⁰⁸
1	78	Belgium	R	-	X	> 8	X	-	-	-	X	X	X	X	Slootmaekers et al. ¹⁰⁹
1	11	China	L	-	X	> 8	X	X	-	-	X	X	-	X	Yuan et al. ¹¹⁰
14	8-9	China	E	-	X	> 20	X	-	-	-	X	-	X	X	Su et al. ¹¹¹

2	265 & 184	Brazil	E	-	X	?	X	-	-	-	-	-	-	-	Ferreira et al. ¹¹²
1	20	Argentina	E	-	X	> 8	X	-	-	-	X	-	-	-	Arias et al. ¹¹³
22	-	Germany	R & L	-	X	> 20	X	X	X	-	-	-	-	X	Roch et al. ⁸¹
1	180	Australia	W	-	X	> 20	X	X	-	-	-	-	X	X	Su et al. ¹¹⁴

332 No.=number, sp.=species, R=river, L=lake, E=estuary, W=wetland, FR=fragments, FI=fibers, AP=anthropogenic particles excluding plastic particles,

333 PL=plastics, Hyd.=hydroxides, Enz.=enzymes, Ox.=oxidizers, Rn=Raman spectroscopy, FTIR=Fourier transform infrared spectroscopy.

334 *Visual examination after the digestion process. In this table, when the minimum size of the MP was not mentioned in the study, the porosity of the

335 filters was considered. †No results for those blanks were provided.

337 Table 2. Summary of plastic characteristics from gut contents of freshwater fish.

Species (n)	Location	Plastic concentration or occurrence			Mean plastic size or range (mm)	Spectroscopic identification?	Reference
		Item/individual	Item/g GC	Contaminated individuals (%)			
<i>Gobio gobio</i> (78)	Several rivers, Belgium	-	-	9	0.67	Yes	109
<i>Gobio gobio</i> (186)	Several rivers, France	-	-	12	-	No	98
<i>Squalius cephalus</i>	Seine River, France	0.16*	0.16*	15	2.67*	Yes	57
22 species	Lakes & rivers, Germany	0.2	-	18.8	0.889	No	81
<i>Alburnus alburnus</i> <i>Perca fluviatilis</i> <i>Rutilus rutilus</i> (10) <i>Leuciscus leuciscus</i>	Lake Geneva, Switzerland	3.1 0 0 0.3	-	7.5	-	No	100
<i>Platichthys flesus</i> <i>Osmerus eperlanus</i>	Thames Estuary, England	-	-	85 20	-	Yes	104
<i>Rutilus rutilus</i> (64)	River Thames, England	0.69	-	32.8	-	Yes	67
<i>Platycephalus indicus</i> <i>Saurida tumbil</i> <i>Sillago sihama</i> <i>Cynoglossus abbreviatus</i>	Musa Estuary, Iran	2.3 2.8 1.5 2.9	-	-	-	No, SEM/EDS	106
<i>Lates niloticus</i> <i>Oreochromis niloticus</i>	Lake Victoria, Tanzania	-	-	20 20	-	Yes	99
<i>Lepomis macrochirus</i> (318) <i>Lepomis megalotis</i> (118)	Brazos River, Texas, U.S.A.	-	-	45	-	No	103
44 species	Several rivers, Texas, U.S.A.	-	-	8.2	-	Yes	101
17 species	Several rivers, U.S.A.	~13	-	85	<1.5	Yes	91
<i>Esox lucius</i> <i>Catostomus commersoni</i> <i>Notropis atheirnoides</i> <i>Pimephales promelas</i> (34)	Wascana Creek, Canada	-	-	73.5	-	No	96

<i>Eucalia inconstans</i>							
11 species	Rio de La Plata, Argentina	19.2	-	100	0.06-4.7	No	56
<i>Micropogonias furnieri</i>	Bahía Blanca Estuary, Argentina	12.1	-	-	2.4 (median)	No	113
16 species	Xingu River, Brazil	-	-	26.7	1-15	Yes	108
46 species	Amazon Estuary, Brazil	1.2	-	13.7	1.82	Yes	52
<i>Hoplosternum littorale</i>	Pajeú River, Brazil	3.6	-	83	<1-12	No	105
69 species	Paraíba Estuary Mamanguape Estuary, Brazil	0.07 0.12	-	9	-	No	52
<i>Cathorops spixii</i> <i>Cathorops agassizii</i> <i>Sciades herzbergii</i>	Goiana Estuary, Brazil	-	-	18 33 17	-	No	97
<i>Stellifer brasiliensis</i> (330) <i>Stellifer stellifer</i>	Goiana Estuary, Brazil	-	-	6.9 9.2	-	No	49
<i>Eugerres brasilianus</i> (240) <i>Eucinostomus melanopterus</i> <i>Diapterus rhombeus</i> (44)	Goiana Estuary, Brazil	-	-	16.2 9.2 11.4	1-5	No	50
<i>Cynoscion acoupa</i>	Goiana Estuary, Brazil	-	-	64.2	<5	No	102
<i>Centropomus undecimalis</i> <i>Centropomus mexicanus</i>	Goiana Estuary, Brazil	1.5 1.4	-	-	~1.25	No	112
6 species	Taihu Lake, China	2.4	3.4	95.7	0.4-24.8	Yes	44
14 species	Yangtze Estuary Hangzhou Bay, China	0.3-4.5 0.5-5.3	0.3-6.2 0.1-8.8	-	22-100 25-100	Yes	111
13 species	Xiangxi River, China	-	-	25.7	0.3-1.8	Yes	59
<i>Gymnocypris przewalskii</i>	Qinghai Lake, China	5.4	-	-	-	Yes	107

<i>Carassius auratus (11)</i>	Poyang Lake, China	-	-	91	0.1-1	Yes	110
<i>Gambusia holbrooki</i>	Melbourne Area, Australia	0.60	-	19.4	0.09-4.86	Yes	114

338 GC=gut contents. The bolded items are the species studied in at least two publications with the number of individuals sampled. *Personal unpublished
 339 data and the data published Collard et al.⁵⁷, which concerned all anthropogenic particles. When the global mean is not provided, we gave the total range.

340 3. Factors influencing MP ingestion by freshwater fish

341

342 Here, we aimed to discuss factors that are likely to influence MP ingestion by freshwater fish foraging
343 for prey. Those factors include specific traits, such as the feeding strategy, and abiotic parameters,
344 such as the environmental MP levels. However, the results in the discussed in situ studies are also
345 influenced by the retention time of MP inside the gut, which is not well understood. The results might
346 be different depending on the size of plastic pieces or the species. Also, the gut content reflects only
347 the instantaneous diet, that is why repeated studies should be performed.

348

349 3.1. Levels in abiotic compartments

350

351 According to studies, several factors are at play when plastic is ingested by fish. The first factor to
352 consider is probably the exposure. Very few studies have investigated both the abiotic contamination
353 and MP ingestion by freshwater fish. Therefore, this section will give an overview of the
354 contamination levels in those matrices as a first step in determining what induces or prevents MP
355 ingestion.

356 Many different units are used for both water and sediment concentrations (Table S1), which is also
357 the case for marine environments.^{115,116} Regarding water contamination, the results are often given in
358 items per cubic meter of water, and the format “items/volume” is the most commonly used. Regarding
359 the sediment, the results are usually expressed in items per kilogram of sediment (dry weight).
360 Remarkably, all except one study chose to express their results per dry weight. We thus encourage
361 researchers to further express sediment concentration per dry weight too. In addition, some authors
362 calculated the mean concentration while others chose to only show a range. Some authors also
363 excluded values equal to zero from the mean calculation, and others did not give any concentration
364 mean or range. While a global mean value may be very restrictive and not representative of all the
365 parameters such as the different rivers sampled or difference in the sampling locations (close or far

366 from an outlet, close or far from urban activities, etc.), we recommend showing as many values as
367 possible. This will allow further studies to choose the most convenient and adapted value to be
368 compared to.

369 When considering studies that performed analyses in both water and sediment samples obtained from
370 the same place, several trends and questions arise. Regarding the abundances of MP, some studies
371 found different patterns for the two compartments.^{59,110,117,118} The authors of these studies discussed
372 those results by mentioning potential factors that could influence MP distribution. MP levels in lake
373 sediments are related to their distance from MP sources.^{119,120} Yuan et al.¹¹⁰ confirmed this fact. The
374 northern and middle regions of Poyang Lake are subject to anthropogenic activities and are the most
375 contaminated parts with regards to the sediment and surface waters, respectively. Sediment and water
376 obtained from the south of Poyang Lake, where less human activities occur, are less contaminated
377 than those of the northern and middle regions. Furthermore, the presence of a wastewater treatment
378 plant in the vicinity of a sampling station might induce higher concentrations of polyethylene
379 terephthalate (PET) fibers.^{117,121}

380 The topography of the environment may also be a factor of the concentration or dilution of MP in
381 sediment.¹²² In addition, the hydrodynamics, which are influenced by the shoreline morphology, may
382 favor or prevent the deposition of plastics in the sediment; a low velocity environment, such as a
383 harbor will enable plastics to settle.^{59,97,120,123}

384 Surprisingly, MP contamination in fish shows different patterns than MP contamination in water or
385 sediment for at least one characteristic (Table S2). Sometimes, a correlation between the water or
386 sediment contamination and fish contamination exists for only one or two parameter(s) (*e.g.*, the
387 shape or the size range) or at one sampling location.⁹¹ Therefore, this shows evidences that exposure
388 is not the only factor involved,⁶⁷ and perhaps, that exposure is not a factor at all.¹²⁴ Of course, MP
389 ingestion cannot occur if there is not any plastic in the environment, but Kim et al.¹²⁴ found that the
390 food concentration is surprisingly more important than the MP concentration in influencing MP
391 ingestion. Kim et al.¹²⁴ exposed zebrafish to various mixtures of MP and food and concluded that MP

392 ingestion increased with increasing concentrations of food. However, in that study, the authors
393 exposed fish to virgin plastic. It is now known that animals react differently to virgin or biofouled
394 plastic. The copepods *Calanus finmarchicus* and female copepods *Acartia longiremis* ingested
395 significantly more biofouled PS beads than virgin PS beads.¹²⁵ Similarly, procellariiform seabirds can
396 be fooled by the dimethyl sulfide signature present on marine plastic particles.¹²⁶ In 2017, it has also
397 been shown that *Engraulis mordax* anchovies responded to medium and high concentrations of
398 biofouled plastic odors with foraging behaviors.¹²⁷ Those behaviors did not appear when these
399 anchovies were exposed to clean plastic particles.

400 Indirectly, surface runoff and seasons also indirectly act on the exposure level.^{112,128} Surface runoff
401 increases the exposure to plastic debris.¹¹² Consequently, the exposure plays a role, but the ingestion
402 also depends on other parameters, such as the age of the plastic particle or the time of the year.

403

404 3.2. Living habitat

405

406 Sediment is a sink for microplastics,^{4,129} and therefore, demersal and benthic fish could be exposed
407 to more MP than pelagic species. Some studies have focused on both pelagic and benthic or demersal
408 fish with varying results. In China, demersal species ingested more plastics than pelagic species.⁴⁴
409 The same assumption was made by McGoran et al.¹⁰⁴; a demersal species, *Platichthys flesus* flounder
410 had ingested far more MP than a pelagic species, *Osmerus eperlanus* smelt. McGoran et al.¹⁰⁴
411 assumed that *P. flesus* had ingested more MP, because it ingested sediment, likely leading to the
412 unintended ingestion of MP. In the marine Scottish waters, demersal fish species have ingested more
413 plastics than pelagic species.⁴⁵ They also found a significant difference between coastal and offshore
414 species, in which the former are more contaminated than the latter. Nevertheless, this latter category
415 was only represented by demersal species, which may lead to the wrong conclusions about the
416 involvement of the fish species coastal habitat on MP ingestion. Not all studies focusing on MP
417 ingestion by pelagic and demersal fish found a difference between these two categories.¹³⁰ Moreover,

418 Rummel et al.²² showed that pelagic feeders had more MP in their stomach than demersal fish. Again,
419 as methods vary from one study to another and as MP ingestion depends on several factors, the results
420 may vary when looking at only one factor.

421

422 3.3. Feeding strategy

423

424 A more commonly discussed parameter is the feeding strategy of fish and their trophic position. It is
425 thought that (top-)predator fish from both fresh and marine environments might be more at risk than
426 organisms at lower trophic levels because of high energy requirements and trophic
427 transfer.^{91,96,102,112,131} Other predators such as marine mammals have ingested MP that are too small
428 to be preyed upon by such larger animals.^{13,132,133} This suggests that secondary ingestion has occurred
429 and thus, MP have accumulated through the food chain. The developmental stage of fish might also
430 influence MP ingestion.⁶⁷ Indeed, many fish change their diet throughout their development.¹³⁴⁻¹³⁷
431 Ferreira et al.¹⁰² studied MP contamination in several ontogenetic phases of *Cynoscion acoupa*. The
432 adults, which are mainly piscivorous, were more contaminated than juveniles and subadults, which
433 exhibit an omnivorous diet. This result leads to the same conclusion previously reported: predator or
434 piscivorous fish are more at risk than fish displaying another type of feeding strategy. Later, Ferreira
435 et al.¹¹² made the same conclusion for two other species (the snook *Centropomus undecimalis* and
436 *Centropomus mexicanus*) obtained from the same location, the Goiana Estuary. Three ontogenetic
437 phases were sampled and compared. Juvenile snook registered the lowest MP ingestion level while
438 the piscivorous adults, registered the highest. Moreover, peaks of MP ingestion by adults coincide
439 with peak of fish ingestion. Other studies did not find such differences,¹⁰⁸ indicating the possibility
440 for other parameters to be involved. For example, in the marine environment, debris categories were
441 related to the feeding behavior of the sampled fish.¹³⁸ Opportunistic feeders ingested all debris
442 categories (i.e., metals, wood, and plastics), while pelagic feeders had only swallowed plastic bags,
443 benthic fish, and hard plastic pieces.

444 Based on the fish feeding habits, the visual characteristics of plastic pieces may be involved in MP
445 ingestion by particulate-feeders. In contrast to ram-feeding, which consists of passively filtering water
446 and is nonselective, particulate-feeding is a selective mode, where prey are visually detected before
447 their capture.¹³⁹ Then, ram-feeders are less likely to be influenced by the visual characteristics of
448 plastics.⁷⁷ Yuan et al.¹¹⁰ noticed that white MP were more frequently found in fish stomachs than in
449 both water and sediment. Yuan et al.¹¹⁰ supposed that *Carassius auratus* could mistake MP for
450 planktonic food. Ory et al.¹⁴⁰ studied *Serirolella violacea* juveniles in controlled conditions and
451 exposed them to food-shaped and food-sized polyamide particles with different colors. As expected,
452 the juveniles preferentially ingested black particles, which is the same color as food pellets. The other
453 colored particles were supposed to be cocaptured with the food, as they were ingested together with
454 the food pellets only. Nevertheless, attention should be paid to the digesting agent used in the
455 extraction process as some agents, such as NaClO, change or discolor the particles.⁶⁸ In a more global
456 framework, the likelihood that beached macroplastics were previously bitten by fish and other
457 organisms depends on the plastic colors.¹⁴¹ Blue and yellow macroplastics were preferentially more
458 than were other colors. Similarly, plastic threads were the main plastic type ingested by *Cynoscion*
459 *acoupa* in the Goiana Estuary, while plastic threads represented only 1.4% of all the plastic debris in
460 the water column.^{102,142} Several colors of threads were ingested, suggesting that the plastic shape was
461 more important than the color for this species. The same observation was made by Carson,¹⁴¹ who
462 found that bottle shaped macroplastics displayed more bite marks than other shapes. Visual and
463 chemical cues thus interfere together with the feeding behavior of fish, making the understanding of
464 MP ingestion in fish even more challenging. Further studies that will consider several parameters are
465 needed to understand the different pathways involved. Increasing the knowledge of those factors will
466 help to suggest and define species for the monitoring of plastic pollution in both fresh and marine
467 waters.

468

469 4. Freshwater fish for the biomonitoring of MP contamination?

470

471 The occurrence of a contaminant in an ecosystem does not mean that the contaminant will cause harm
472 to the populations inhabiting the ecosystem. Biota is needed to link the pollutant levels in abiotic
473 compartments and in organisms' tissue to potential adverse effects.¹⁴³ Using fish species for the
474 assessment of the quality of aquatic ecosystems is highly relevant. Fish are found everywhere in
475 aquatic environments and have an intermediate trophic position, linking the lower trophic positions
476 to the higher trophic positions (Beyer 1996 in van der Oost et al.¹⁴³). Nevertheless, attention must be
477 paid when choosing a species for monitoring. The GESAMP⁴⁶ report suggested a list of criteria that
478 helps to define good species for monitoring. This species, or group of species, must be representative
479 of different life histories, phylogenies, sizes, ages and developmental stages. General strategies are
480 proposed, which include opportunistic sampling, market sampling of commercial species and
481 targeting biota. In this report, it is stated that fish, as targeted species or those purchased from the
482 markets, could constitute a good monitoring group for the ingestion of MP.⁴⁶

483 Given the numerous factors influencing the ingestion of MP by fish, choosing only one indicator
484 species or a group of species is challenging and perhaps impossible. The ecology and feeding habits
485 of the chosen species must be known. It is important to know where the fish, as well as all its
486 developmental stages, is located within the food web, for instance. Here, we present a summary of
487 the parameters to be considered when choosing an indicator fish species, for both fresh and marine
488 environments:

- 489 - *Trophic position.* As discussed in the previous section, predators, especially top-predators, are
490 more likely to ingest MP through food transfer. However, monitoring MP pollution with top-
491 predators will not accurately indicate the contamination level of the environment (water and
492 sediment) but will give global information on the trophic web and the ecosystem.
- 493 - *Feeding strategy.* Ram-feeding fish passively filter water without any visual search for food.
494 They may be good indicators of the MP levels in the surrounding water. In this case, in
495 addition to gut contents, we advise researchers to also examine the gill rakers, which are bony

496 or cartilaginous structures attached to the branchial arches that retain filtered particles inside
497 the mouth. Those analyses will complete the GC results.

498 - *Migrations*. Many fish migrate, either vertically (diel migrations) or horizontally (seasonal
499 migrations), and sometimes fish migrate both ways. Depending on the area chosen to be
500 studied, all types of migration can be valuable. Diel migrations could inform researchers about
501 the MP levels in the pelagic zone, as the surface, seasonal and spatial migrations could provide
502 data for the same species but in different areas. Utilizing the same species is important, as it
503 avoids some bias caused by specific related factors, such as the fish length, the mouth structure
504 and the fish vision. Whatever the chosen species and area, the sampling protocol must be
505 designed considering those migration types. Fish must be sampled at the same time of the day
506 or year if only one location or one water column compartment is targeted.

507 - *Commercial value*. Collecting commercially valued fish is advantageous as this is a cost-
508 effective method for the assessment of human exposure through the chosen species.⁴⁶ Indeed,
509 commercial fish can be purchased at markets, which eliminates evident sampling costs.
510 However, it may not be possible to obtain precise information regarding the sampling location.

511 - *Living habitat*. Benthic and demersal species will provide information about a sea
512 compartment that is different than the sea compartment of pelagic species. Demersal fish will
513 give a less accurate overview of the benthic compartment than benthic fish, as demersal fish
514 are not fully associated with the sediment.

515 - *Distribution*. For comparison and repeatability purposes, the chosen species should have the
516 widest distribution possible or should at least belong to a higher taxon (genus or family) that
517 is distributed worldwide distributed in order to avoid biases linked to the different species
518 used in various studies.

519

520 In summary, the perfect fish species used to monitor MP pollution in fresh and marine water is either
521 a top-predator or a ram-feeding species with a large distribution area and a commercial value. If the

522 species is restricted to a small area, similar species must be found in other areas of the world. The
523 migration of the selected fish must be well known and considered when sampling. Researchers must
524 then carefully choose the correct species according to the information they need.

525

526 5. Perspectives and recommendations

527

528 To provide perspectives for further research, we want to highlight three steps of protocols that are of
529 great importance: the number of samples analyzed, the size limit of the extracted particles and their
530 identification.

531 Sample sizes vary greatly between studies, as sizes vary from 1 to 330 individuals for a single species.

532 A sample size of 50 individuals has been determined to be sufficient for plastic pollution research.⁵³

533 A couple of samples is not useful for research; in addition to being statistically irrelevant, time and/or
534 money can be saved by not sampling such small sample sizes and perhaps used for better purposes.

535 Achieving 50 individuals per studied criterion can be challenging, and therefore, we recommend
536 sampling as close to 50 individuals as possible by sampling the same species from a specific location,
537 for example.

538 Studies have rarely identified the target size range of MP extracted from fish gut contents (Table 2).

539 The only information available is the mesh size of the filters used during the extraction step when
540 present. Usually, that mesh size is approximately a micrometer, which allows researchers to extract
541 very small particles, but such particles are hardly observed visually and analyzed spectroscopically.

542 In contrast, focusing on large MP might lead to an underestimation of the number of ingested
543 particles. Both large and small fish are likely to ingest small MP (< 100 μm) either by transfer from
544 prey to predator or through their feeding behavior. It is thus critical to specify which threshold is
545 applied in each study to promote comparability. This threshold must be defined according to the
546 identification method used; FTIR and Raman spectroscopies allow researchers to analyze MP that
547 are smaller than the particles visible by visual observation through a stereomicroscope.

548 MP have often been identified by visual inspection, although the identification techniques used have
549 seemingly improved in recent studies. Only two studies on MP contamination in freshwater fish
550 included all dyed particles, even if the particles were not made of plastic (Table 1). We believe that
551 it is important to report all types of anthropogenic contamination in fish tissues, especially textile
552 fibers, which are usually dyed and released in large amounts into the environment.^{144,145}

553 With regards to contamination, we suggest that (1) sample collection is performed with as few plastic
554 materials as possible, (2) all solutions, including tap water, must be filtered with filters having the
555 same porosity as the one used to filter the samples and stored in glass jars, (3) plastic materials should
556 be avoided during sample treatment, (4) if the laboratory work cannot be performed in a clean room,
557 the laboratory should then be ventilated as much as possible, (5) all equipment must be covered with
558 clean non-plastic materials when not in use and (6) the laboratory operator should wear nonsynthetic
559 clothes, a cotton lab coat and gloves. Avoiding plastic during the sampling and treatment steps is not
560 easy, especially regarding the textiles worn, solution storage and use of nonsynthetic gloves, but for
561 the latter at least, an FTIR or Raman spectrum can be recorded and matched with the potential
562 contaminating particles found in the samples. If contamination does occur, those particles can then
563 be excluded from the results.

564 As previously recommended by published reviews,^{40,47,48} standardized protocols are required. Thus
565 we briefly recommend an overall optimal methodology that can be used when evaluating MP
566 pollution by working with fish gut samples: (1) the number of sampled individuals should be as close
567 to 50 as possible, (2) a MP target size down to 1 μm should be considered to fit the MP definition,
568 (3) enzymes should be used as digesting agents, (4) identification should be performed by
569 spectroscopy, (5) qualitative observations should be made by light microscopy, (6) as many data as
570 possible (averages and ranges, global contamination or per category, etc.) should be provided, (7) the
571 results should be expressed using several units and per category studied (feeding strategy,
572 developmental stage, etc.), and (8) procedural blank samples should be prepared and analyzed.
573 Spectroscopic analyses can be performed on a subsample of the total amount of extracted particles.

574 However, when focusing on fish contamination, it is uncommon to find tens of particles in a single
575 individual. Therefore, we recommend analyzing all particles extracted from the fish GCs. Visual
576 observations are not recommended for sorting or identifying microplastics but are required to take
577 pictures and to measure the MP found. We encourage researchers to keep that step only for those
578 purposes. The global evaluation of the protocol quality can be made thanks to the quality assessment
579 system proposed by Hermesen et al.⁴⁸ This quality assessment has been adapted for MP ingestion from
580 the CRED scoring system¹⁴⁶ and provides a good overview of the protocol strength.

581

582 This review examines MP in the digestive tracts of freshwater fish, but MP appear to translocate to
583 other organs in several aquatic taxonomic groups, including freshwater fish.^{57,61,111,147-149} The impacts
584 of MP translocation are not well known, and MP pathways still need to be understood. The liver has
585 primarily been studied, but other tissues are worth investigating. Based on the human health
586 perspective, muscle is obviously the first tissue that comes to mind, as muscle is almost the only part
587 of the fish we consume. Few studies have examined MP in fish muscle,^{57,106,111,147} and when MP are
588 found in the muscle, their pathways are unknown. In this regard, in vitro studies using biofouled
589 plastics are needed to provide more information about the translocation process(es).

590 Selecting one or more species suitable for monitoring is currently challenging. First, studies must be
591 performed on MP ingestion by fish and MP contamination in their nearby environment. Studies
592 should be performed at the same time and, of course, by utilizing proper methods with chemical
593 treatment and identification. This will allow scientists to determine which species reflects the best
594 environmental contamination and thus to decide which species can be defined as an indicator.
595 Furthermore, assessing the species monitoring process is only the first step towards obtaining a more
596 global understanding of MP pollution in biota. To date, most studies on MP ingestion by fish are quite
597 descriptive and contain suggested explanations that are usually not demonstrated, and potential
598 impacts are listed. The quantity and quality of research require improvements to be made to assess
599 the ecological risk posed by MP.¹⁵⁰ A description of the contamination levels is obviously of great

600 importance to the scientific community, but we think that only studying MP contamination is not
601 sufficient. Microplastics alone have been found to negatively impact fish,^{25,151,152} but MPs are part of
602 a cocktail of pollutants containing additives, *e.g.*, phthalates and bisphenols, dyes, organic pollutants,
603 among others. These pollutants are also known to affect fish in several ways.¹⁵³⁻¹⁵⁵ Consequently, MP
604 should be monitored concomitantly with other pollutants to more accurately reflect the ecological
605 state of freshwaters with regards to pollution.

606

607 In conclusion, this review aims to highlight the need for the standardization of methodologies and to
608 give an overview of the factors involved in the ingestion of MP by freshwater fish. The first step of
609 standardization can be to follow several recommendations, such as suggesting that a sufficient
610 number of individuals be sampled and recommending the systematic use of spectroscopy to identify
611 polymer particles. With regards to biomonitoring, studies should be performed using the same overall
612 methodology and the same or very similar fish species. These species must be chosen for the purpose
613 of monitoring and based on their biological characteristics: top-predator and ram-filtering species
614 provide useful information with regards to plastic contamination.

615

616 6. Supporting information

617

618 The Supporting Information is available free of charge on the ACS Publications website at DOI:
619 XXX.

620 An overview of the studies reporting plastic contamination in freshwaters and a summary of data
621 regarding MP contamination in abiotic compartments and fish at the same location.

622

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624

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630

631 8. References

632

- 633 (1) Ryan, P. G. *A Brief History of Marine Litter Research*. In: *Marine Anthropogenic Litter*; Bergmann,
634 M., Gutow, L., Klages, M., Eds.; Springer International Publishing: Cham, 2015.
635 <https://doi.org/10.1007/978-3-319-16510-3>.
- 636 (2) Law, K. L.; Morét-Ferguson, S. E.; Goodwin, D. S.; Zettler, E. R.; Deforce, E.; Kukulka, T.;
637 Proskurowski, G. Distribution of Surface Plastic Debris in the Eastern Pacific Ocean from an 11-Year
638 Data Set. *Environ. Sci. Technol.* **2014**, *48* (9), 4732–4738. <https://doi.org/10.1021/es4053076>.
- 639 (3) Gago, J.; Carretero, O.; Filgueiras, A. V.; Viñas, L. Synthetic Microfibers in the Marine
640 Environment: A Review on Their Occurrence in Seawater and Sediments. *Mar. Pollut. Bull.* **2018**,
641 *127*, 365–376. <https://doi.org/10.1016/j.marpolbul.2017.11.070>.
- 642 (4) Woodall, L. C.; Sanchez-Vidal, A.; Canals, M.; Paterson, G. L. J.; Coppock, R.; Sleight, V.; Calafat,
643 A.; Rogers, A. D.; Narayanaswamy, B. E.; Thompson, R. C. The Deep Sea Is a Major Sink for
644 Microplastic Debris. *R. Soc. Open Sci.* **2014**, *1* (4), 140317–140317.
645 <https://doi.org/10.1098/rsos.140317>.
- 646 (5) Tekman, M. B.; Krumpen, T.; Bergmann, M. Marine Litter on Deep Arctic Seafloor Continues to
647 Increase and Spreads to the North at the HAUSGARTEN Observatory. *Deep Sea Res. Part I*
648 *Oceanogr. Res. Pap.* **2017**, *120*, 88–99. <https://doi.org/https://doi.org/10.1016/j.dsr.2016.12.011>.
- 649 (6) Obbard, R. W.; Sadri, S.; Wong, Y. Q.; Khitun, A. A.; Baker, I.; Thompson, R. C. Global Warming
650 Releases Microplastic Legacy Frozen in Arctic Sea Ice. *Earth's Futur.* **2014**, *2* (6), 315–320.
651 <https://doi.org/10.1002/2014EF000240>.
- 652 (7) Peeken, I.; Primpke, S.; Beyer, B.; Gütermann, J.; Katlein, C.; Krumpen, T.; Bergmann, M.;
653 Hehemann, L.; Gerdts, G. Arctic Sea Ice Is an Important Temporal Sink and Means of Transport for
654 Microplastic. *Nat. Commun.* **2018**, *9* (1), 1505. <https://doi.org/10.1038/s41467-018-03825-5>.
- 655 (8) Liu, M.; Lu, S.; Song, Y.; Lei, L.; Hu, J.; Lv, W.; Zhou, W.; Cao, C.; Shi, H.; Yang, X.; et al.
656 Microplastic and Mesoplastic Pollution in Farmland Soils in Suburbs of Shanghai, China. *Environ.*
657 *Pollut.* **2018**, *242*, 855–862. <https://doi.org/10.1016/j.envpol.2018.07.051>.
- 658 (9) Dris, R.; Gasperi, J.; Saad, M.; Mirande, C.; Tassin, B. Synthetic Fibers in Atmospheric Fallout: A
659 Source of Microplastics in the Environment? *Mar. Pollut. Bull.* **2016**, *104* (1–2), 290–293.

- 660 <https://doi.org/10.1016/j.marpolbul.2016.01.006>.
- 661 (10) Cai, L.; Wang, J.; Peng, J.; Tan, Z.; Zhan, Z.; Tan, X.; Chen, Q. Characteristic of Microplastics in the
662 Atmospheric Fallout from Dongguan City, China: Preliminary Research and First Evidence. *Environ.*
663 *Sci. Pollut. Res.* **2017**, *24* (32), 24928–24935. <https://doi.org/10.1007/s11356-017-0116-x>.
- 664 (11) *Marine Anthropogenic Litter*; Bergmann, M., Gutow, L., Klages, M., Eds.; Springer International
665 Publishing: Cham, 2015. <https://doi.org/10.1007/978-3-319-16510-3>.
- 666 (12) Lusher, A. L.; Hernandez-Milian, G.; O'Brien, J.; Berrow, S.; O'Connor, I.; Officer, R. Microplastic
667 and Macroplastic Ingestion by a Deep Diving, Oceanic Cetacean: The True's Beaked Whale
668 *Mesoplodon Mirus*. *Environ. Pollut.* **2015**, *199*, 185–191.
- 669 (13) Nelms, S. E.; Galloway, T. S.; Godley, B. J.; Jarvis, D. S.; Lindeque, P. K. Investigating Microplastic
670 Trophic Transfer in Marine Top Predators. *Environ. Pollut.* **2018**, *238*, 999–1007.
671 <https://doi.org/10.1016/J.ENVPOL.2018.02.016>.
- 672 (14) Zhu, J.; Yu, X.; Zhang, Q.; Li, Y.; Tan, S.; Li, D.; Yang, Z.; Wang, J. Cetaceans and Microplastics:
673 First Report of Microplastic Ingestion by a Coastal Delphinid, *Sousa Chinensis*. *Sci. Total Environ.*
674 **2019**, *659*, 649–654. <https://doi.org/10.1016/j.scitotenv.2018.12.389>.
- 675 (15) Trevail, A. M.; Gabrielsen, G. W.; Kühn, S.; Van Franeker, J. A. Elevated Levels of Ingested Plastic
676 in a High Arctic Seabird, the Northern Fulmar (*Fulmarus Glacialis*). *Polar Biol.* **2015**, *38* (7), 975–
677 981. <https://doi.org/10.1007/s00300-015-1657-4>.
- 678 (16) Zhao, S.; Zhu, L.; Li, D. Microscopic Anthropogenic Litter in Terrestrial Birds from Shanghai,
679 China: Not Only Plastics but Also Natural Fibers. *Sci. Total Environ.* **2016**, *550*, 1110–1115.
680 <https://doi.org/10.1016/j.scitotenv.2016.01.112>.
- 681 (17) Reynolds, C.; Ryan, P. G. Micro-Plastic Ingestion by Waterbirds from Contaminated Wetlands in
682 South Africa. *Mar. Pollut. Bull.* **2018**, *126*, 330–333.
683 <https://doi.org/10.1016/J.MARPOLBUL.2017.11.021>.
- 684 (18) Devriese, L. I.; van der Meulen, M. D.; Maes, T.; Bekaert, K.; Paul-Pont, I.; Frère, L.; Robbens, J.;
685 Vethaak, A. D. Microplastic Contamination in Brown Shrimp (*Crangon Crangon*, Linnaeus 1758)
686 from Coastal Waters of the Southern North Sea and Channel Area. *Mar. Pollut. Bull.* **2015**, *98* (1),
687 179–187. <https://doi.org/10.1016/j.marpolbul.2015.06.051>.
- 688 (19) Iannilli, V.; Di Gennaro, A.; Lecce, F.; Sighicelli, M.; Falconieri, M.; Pietrelli, L.; Poeta, G.; Battisti,

- 689 C. Microplastics in *Talitrus Saltator* (Crustacea, Amphipoda): New Evidence of Ingestion from
690 Natural Contexts. *Environ. Sci. Pollut. Res.* **2018**, *25* (28), 28725–28729.
691 <https://doi.org/10.1007/s11356-018-2932-z>.
- 692 (20) Hurley, R. R.; Woodward, J. C.; Rothwell, J. J. Ingestion of Microplastics by Freshwater Tubifex
693 Worms. *Environ. Sci. Technol.* **2017**, *51* (21), 12844–12851. <https://doi.org/10.1021/acs.est.7b03567>.
- 694 (21) Bråte, I. L. N.; Eidsvoll, D. P.; Steindal, C. C.; Thomas, K. V. Plastic Ingestion by Atlantic Cod (
695 *Gadus Morhua*) from the Norwegian Coast. *Mar. Pollut. Bull.* **2016**, *112* (1–2), 105–110.
696 <https://doi.org/10.1016/j.marpolbul.2016.08.034>.
- 697 (22) Rummel, C. D.; Löder, M. G. J.; Fricke, N. F.; Lang, T.; Griebeler, E.-M.; Janke, M.; Gerdts, G.
698 Plastic Ingestion by Pelagic and Demersal Fish from the North Sea and Baltic Sea. *Mar. Pollut. Bull.*
699 **2016**, *102* (1), 134–141. <https://doi.org/10.1016/j.marpolbul.2015.11.043>.
- 700 (23) Collicutt, B.; Juanes, F.; Dudas, S. E. Microplastics in Juvenile Chinook Salmon and Their Nearshore
701 Environments on the East Coast of Vancouver Island. *Environ. Pollut.* **2019**, *244*, 135–142.
702 <https://doi.org/10.1016/j.envpol.2018.09.137>.
- 703 (24) Arthur, C.; Baker, J.; Bamford, H. Proceedings of the International Research Workshop on the
704 Occurrence, Effects and Fate of Microplastic Marine Debris. In *Proceedings of the International*
705 *Research Workshop on the Occurrence, Effects and Fate of Microplastic Marine Debris.*; Technical
706 Memorandum NOS-OR&R-30; Proceedings of the International Research Workshop on the
707 Occurrence, Effects, and Fate of Microplastic Marine Debris Courtney Arthur, Joel Baker, and
708 Holly Bamford (eds.) National Oceanic and Atmospheric Administration: University of Washington
709 Tacoma campus in Tacoma, Washington, USA, 2008.
- 710 (25) Anbumani, S.; Kakkar, P. Ecotoxicological Effects of Microplastics on Biota: A Review. *Environ.*
711 *Sci. Pollut. Res.* **2018**, *25* (15), 14373–14396. <https://doi.org/10.1007/s11356-018-1999-x>.
- 712 (26) Barboza, L. G. A.; Dick Vethaak, A.; Lavorante, B. R. B. O.; Lundebye, A.-K.; Guilhermino, L.
713 Marine Microplastic Debris: An Emerging Issue for Food Security, Food Safety and Human Health.
714 *Mar. Pollut. Bull.* **2018**, *133*, 336–348. <https://doi.org/10.1016/j.marpolbul.2018.05.047>.
- 715 (27) Choi, J. S.; Jung, Y.-J.; Hong, N.-H.; Hong, S. H.; Park, J.-W. Toxicological Effects of Irregularly
716 Shaped and Spherical Microplastics in a Marine Teleost, the Sheepshead Minnow (*Cyprinodon*
717 *Variegatus*). *Mar. Pollut. Bull.* **2018**, *129* (1), 231–240.

- 718 <https://doi.org/10.1016/j.marpolbul.2018.02.039>.
- 719 (28) de Sá, L. C.; Luís, L. G.; Guilhermino, L. Effects of Microplastics on Juveniles of the Common Goby
720 (Pomatoschistus Microps): Confusion with Prey, Reduction of the Predatory Performance and
721 Efficiency, and Possible Influence of Developmental Conditions. *Environ. Pollut.* **2015**, *196*, 359–
722 362. <https://doi.org/10.1016/j.envpol.2014.10.026>.
- 723 (29) Small, C.; Nicholls, R. J. A Global Analysis of Human Settlement in Coastal Zones. *J. Coast. Res.*
724 **2003**, *19* (3), 584–599.
- 725 (30) Kummu, M.; de Moel, H.; Ward, P. J.; Varis, O. How Close Do We Live to Water? A Global
726 Analysis of Population Distance to Freshwater Bodies. *PLoS One* **2011**, *6* (6), e20578.
727 <https://doi.org/10.1371/journal.pone.0020578>.
- 728 (31) Lebreton, L. C. M.; van der Zwet, J.; Damsteeg, J.-W.; Slat, B.; Andrady, A.; Reisser, J. River Plastic
729 Emissions to the World's Oceans. *Nat. Commun.* **2017**, *8*, 15611.
730 <https://doi.org/10.1038/ncomms15611>.
- 731 (32) Carr, S. A.; Liu, J.; Tesoro, A. G. Transport and Fate of Microplastic Particles in Wastewater
732 Treatment Plants. *Water Res.* **2016**, *91*, 174–182. <https://doi.org/10.1016/j.watres.2016.01.002>.
- 733 (33) Mason, S. A.; Garneau, D.; Sutton, R.; Chu, Y.; Ehmann, K.; Barnes, J.; Fink, P.; Papazissimos, D.;
734 Rogers, D. L. Microplastic Pollution Is Widely Detected in US Municipal Wastewater Treatment
735 Plant Effluent. *Environ. Pollut.* **2016**, *218*, 1045–1054. <https://doi.org/10.1016/j.envpol.2016.08.056>.
- 736 (34) Free, C. M.; Jensen, O. P.; Mason, S. A.; Eriksen, M.; Williamson, N. J.; Boldgiv, B. High-Levels of
737 Microplastic Pollution in a Large, Remote, Mountain Lake. *Mar. Pollut. Bull.* **2014**, *85* (1), 156–163.
738 <https://doi.org/10.1016/j.marpolbul.2014.06.001>.
- 739 (35) Baldwin, A. K.; Corsi, S. R.; Mason, S. A. Plastic Debris in 29 Great Lakes Tributaries: Relations to
740 Watershed Attributes and Hydrology. *Environ. Sci. Technol.* **2016**, *50* (19), 10377–10385.
741 <https://doi.org/10.1021/acs.est.6b02917>.
- 742 (36) Eriksen, M.; Maximenko, N.; Thiel, M.; Cummins, A.; Lattin, G.; Wilson, S.; Hafner, J.; Zellers, A.;
743 Rifman, S. Plastic Pollution in the South Pacific Subtropical Gyre. *Mar. Pollut. Bull.* **2013**, *68* (1–2),
744 71–76.
- 745 (37) Gasperi, J.; Dris, R.; Bonin, T.; Rocher, V.; Tassin, B. Assessment of Floating Plastic Debris in
746 Surface Water along the Seine River. *Environ. Pollut.* **2014**, *195*, 163–166.

- 747 <https://doi.org/10.1016/j.envpol.2014.09.001>.
- 748 (38) Lechner, A.; Keckeis, H.; Lumesberger-Loisl, F.; Zens, B.; Krusch, R.; Tritthart, M.; Glas, M.;
749 Schludermann, E. The Danube so Colourful: A Potpourri of Plastic Litter Outnumbers Fish Larvae in
750 Europe's Second Largest River. *Environ. Pollut.* **2014**, *188*, 177–181.
751 <https://doi.org/http://dx.doi.org/10.1016/j.envpol.2014.02.006>.
- 752 (39) Dris, R.; Imhof, H.; Sanchez, W.; Gasperi, J.; Galgani, F.; Tassin, B.; Laforsch, C. Beyond the Ocean:
753 Contamination of Freshwater Ecosystems with (Micro-)Plastic Particles. *Environ. Chem.* **2015**, *12*
754 (5), 539–550. <https://doi.org/10.1071/EN14172>.
- 755 (40) O'Connor, J. D.; Mahon, A. M.; Ramsperger, A. F. R. M.; Trotter, B.; Redondo-Hasselerharm, P. E.;
756 Koelmans, A. A.; Lally, H. T.; Murphy, S. Microplastics in Freshwater Biota: A Critical Review of
757 Isolation, Characterization, and Assessment Methods. *Glob. Challenges* **2019**, 1800118.
758 <https://doi.org/10.1002/gch2.201800118>.
- 759 (41) Powers, D. Fish as Model Systems. *Science (80-.)*. **1989**, *246* (4928), 352–358.
760 <https://doi.org/10.1126/science.2678474>.
- 761 (42) Nelson, J. S. *Fishes of the World*; John Wiley & Sons, Inc.: Hoboken, 2006.
- 762 (43) Markic, A.; Gaertner, J.-C.; Gaertner-Mazouni, N.; Koelmans, A. A. Plastic Ingestion by Marine Fish
763 in the Wild. *Crit. Rev. Environ. Sci. Technol.* **2019**, 1–41.
764 <https://doi.org/10.1080/10643389.2019.1631990>.
- 765 (44) Jabeen, K.; Su, L.; Li, J.; Yang, D.; Tong, C.; Mu, J.; Shi, H. Microplastics and Mesoplastics in Fish
766 from Coastal and Fresh Waters of China. *Environ. Pollut.* **2017**, *221*, 141–149.
767 <https://doi.org/http://dx.doi.org/10.1016/j.envpol.2016.11.055>.
- 768 (45) Murphy, F.; Russell, M.; Ewins, C.; Quinn, B. The Uptake of Macroplastic & Microplastic by
769 Demersal & Pelagic Fish in the Northeast Atlantic around Scotland. *Mar. Pollut. Bull.* **2017**, *122* (1–
770 2), 353–359. <https://doi.org/10.1016/j.marpolbul.2017.06.073>.
- 771 (46) GESAMP. *Guidelines or the Monitoring and Assessment of Plastic Litter and Microplastics in the*
772 *Ocean*; Kershaw, P. J., Turra, A., Galgani, F., Eds.; 2019.
- 773 (47) Lusher, A. L.; Welden, N. A.; Sobral, P.; Cole, M. Sampling, Isolating and Identifying Microplastics
774 Ingested by Fish and Invertebrates. *Anal. Methods* **2017**, *9* (9), 1346–1360.
775 <https://doi.org/10.1039/C6AY02415G>.

- 776 (48) Hermsen, E.; Mintenig, S. M.; Besseling, E.; Koelmans, A. A. Quality Criteria for the Analysis of
777 Microplastic in Biota Samples: A Critical Review. *Environ. Sci. Technol.* **2018**, *52* (18), 10230–
778 10240. <https://doi.org/10.1021/acs.est.8b01611>.
- 779 (49) Dantas, D. V.; Barletta, M.; da Costa, M. F. The Seasonal and Spatial Patterns of Ingestion of
780 Polyfilament Nylon Fragments by Estuarine Drums (Sciaenidae). *Environ. Sci. Pollut. Res.* **2012**, *19*
781 (2), 600–606.
- 782 (50) Ramos, J.; Barletta, M.; Costa, M. Ingestion of Nylon Threads by Gerreidae While Using a Tropical
783 Estuary as Foraging Grounds. *Aquat. Biol.* **2012**, *17* (1), 29–34. <https://doi.org/10.3354/ab00461>.
- 784 (51) Vendel, A. L.; Bessa, F.; Alves, V. E. N.; Amorim, A. L. A.; Patrício, J.; Palma, A. R. T. Widespread
785 Microplastic Ingestion by Fish Assemblages in Tropical Estuaries Subjected to Anthropogenic
786 Pressures. *Mar. Pollut. Bull.* **2017**, *117* (1–2), 448–455.
787 <https://doi.org/10.1016/j.marpolbul.2017.01.081>.
- 788 (52) Pegado, T. de S. e S.; Schmid, K.; Winemiller, K. O.; Chelazzi, D.; Cincinelli, A.; Dei, L.; Giarrizzo,
789 T. First Evidence of Microplastic Ingestion by Fishes from the Amazon River Estuary. *Mar. Pollut.*
790 *Bull.* **2018**, *133*, 814–821. <https://doi.org/10.1016/J.MARPOLBUL.2018.06.035>.
- 791 (53) Galgani, F.; Hanke, G.; Werner, S.; Oosterbaan, L.; Nilsson, P.; Fleet, D.; Kinsey, S.; Thompson, R.
792 C.; van Franeker, J.; Vlachogianna, T.; et al. *Guidance on Monitoring of Marine Litter in European*
793 *Seas*; 2013.
- 794 (54) ICES. *OSPAR Request on Development of a Common Monitoring Protocol for Plastic Particles in*
795 *Fish Stomachs and Selected Shellfish on the Basis of Existing Fish Disease Surveys*; 2015.
- 796 (55) Van Franeker, J.; Meijboom, A. *Litter NSV, Marine Litter Monitoring by Northern Fulmars; a Pilot*
797 *Study*; Wageningen, 2002.
- 798 (56) Pazos, R. S.; Maiztegui, T.; Colautti, D. C.; Paracampo, A. H.; Gómez, N. Microplastics in Gut
799 Contents of Coastal Freshwater Fish from Río de La Plata Estuary. *Mar. Pollut. Bull.* **2017**, *122* (1–
800 2), 85–90. <https://doi.org/10.1016/j.marpolbul.2017.06.007>.
- 801 (57) Collard, F.; Gasperi, J.; Gilbert, B.; Eppe, G.; Azimi, S.; Rocher, V.; Tassin, B. Anthropogenic
802 Particles in the Stomach Contents and Liver of the Freshwater Fish *Squalius Cephalus*. *Sci. Total*
803 *Environ.* **2018**, *643*. <https://doi.org/10.1016/j.scitotenv.2018.06.313>.
- 804 (58) Dehaut, A.; Cassone, A.-L.; Frère, L.; Hermabessiere, L.; Himber, C.; Rinnert, E.; Rivière, G.;

- 805 Lambert, C.; Soudant, P.; Huvet, A.; et al. Microplastics in Seafood: Benchmark Protocol for Their
806 Extraction and Characterization. *Environ. Pollut.* **2016**, *215*, 223–233.
807 <https://doi.org/10.1016/j.envpol.2016.05.018>.
- 808 (59) Zhang, K.; Xiong, X.; Hu, H.; Wu, C.; Bi, Y.; Wu, Y.; Zhou, B.; Lam, P. K. S.; Liu, J. Occurrence
809 and Characteristics of Microplastic Pollution in Xiangxi Bay of Three Gorges Reservoir, China.
810 *Environ. Sci. Technol.* **2017**, *51* (7), 3794–3801. <https://doi.org/10.1021/acs.est.7b00369>.
- 811 (60) Claessens, M.; Van Cauwenberghe, L.; Vandegehuchte, M. B.; Janssen, C. R. New Techniques for
812 the Detection of Microplastics in Sediments and Field Collected Organisms. *Mar. Pollut. Bull.* **2013**,
813 *70* (1–2), 227–233.
- 814 (61) Avio, C. G.; Gorbi, S.; Regoli, F. Experimental Development of a New Protocol for Extraction and
815 Characterization of Microplastics in Fish Tissues: First Observations in Commercial Species from
816 Adriatic Sea. *Mar. Environ. Res.* **2015**, *111*, 18–26. <https://doi.org/10.1016/j.marenvres.2015.06.014>.
- 817 (62) Catarino, A. I.; Thompson, R.; Sanderson, W.; Henry, T. B. Development and Optimization of a
818 Standard Method for Extraction of Microplastics in Mussels by Enzyme Digestion of Soft Tissues.
819 *Environ. Toxicol. Chem.* **2017**, *36* (4), 947–951. <https://doi.org/10.1002/etc.3608>.
- 820 (63) Cole, M.; Webb, H.; Lindeque, P. K.; Fileman, E. S.; Halsband, C.; Galloway, T. S. Isolation of
821 Microplastics in Biota-Rich Seawater Samples and Marine Organisms. *Sci. Rep.* **2014**, *4*, 4528.
822 <https://doi.org/10.1038/srep04528>.
- 823 (64) Railo, S.; Talvitie, J.; Setälä, O.; Koistinen, A.; Lehtiniemi, M. Application of an Enzyme Digestion
824 Method Reveals Microlitter in *Mytilus Trossulus* at a Wastewater Discharge Area. *Mar. Pollut. Bull.*
825 **2018**, *130*, 206–214. <https://doi.org/10.1016/j.marpolbul.2018.03.022>.
- 826 (65) von Friesen, L. W.; Granberg, M. E.; Hassellöv, M.; Gabrielsen, G. W.; Magnusson, K. An Efficient
827 and Gentle Enzymatic Digestion Protocol for the Extraction of Microplastics from Bivalve Tissue.
828 *Mar. Pollut. Bull.* **2019**, *142*, 129–134. <https://doi.org/10.1016/j.marpolbul.2019.03.016>.
- 829 (66) Tagg, A. S.; Harrison, J. P.; Ju-Nam, Y.; Sapp, M.; Bradley, E. L.; Sinclair, C. J.; Ojeda, J. J.
830 Fenton's Reagent for the Rapid and Efficient Isolation of Microplastics from Wastewater. *Chem.*
831 *Commun.* **2017**, *53* (2), 372–375. <https://doi.org/10.1039/C6CC08798A>.
- 832 (67) Horton, A. A.; Jürgens, M. D.; Lahive, E.; van Bodegom, P. M.; Vijver, M. G. The Influence of
833 Exposure and Physiology on Microplastic Ingestion by the Freshwater Fish *Rutilus Rutilus* (Roach)

- 834 in the River Thames, UK. *Environ. Pollut.* **2018**, *236*, 188–194.
835 <https://doi.org/10.1016/J.ENVPOL.2018.01.044>.
- 836 (68) Collard, F.; Gilbert, B.; Eppe, G.; Parmentier, E.; Das, K. Detection of Anthropogenic Particles in
837 Fish Stomachs: An Isolation Method Adapted to Identification by Raman Spectroscopy. *Arch.*
838 *Environ. Contam. Toxicol.* **2015**, *69* (3), 331–339. <https://doi.org/10.1007/s00244-015-0221-0>.
- 839 (69) Kühn, S.; van Werven, B.; van Oyen, A.; Meijboom, A.; Bravo Rebolledo, E. L.; van Franeker, J. A.
840 The Use of Potassium Hydroxide (KOH) Solution as a Suitable Approach to Isolate Plastics Ingested
841 by Marine Organisms. *Mar. Pollut. Bull.* **2017**, *115* (1–2), 86–90.
842 <https://doi.org/10.1016/j.marpolbul.2016.11.034>.
- 843 (70) Barrows, A. P. W.; Cathey, S. E.; Petersen, C. W. Marine Environment Microfiber Contamination:
844 Global Patterns and the Diversity of Microparticle Origins. *Environ. Pollut.* **2018**, *237*, 275–284.
845 <https://doi.org/10.1016/J.ENVPOL.2018.02.062>.
- 846 (71) Darbra, R. M.; Gonzalez Dan, J. R.; Casal, J.; Agueda, A.; Capri, E.; Fait, G.; Schuhmacher, M.;
847 Nadal, M.; Rovira, J.; Grundmann, V.; et al. Additives in the Textile Industry. In *Global Risk-Based*
848 *Management of Chemical Additives I. Production, usage, and environmental occurrence*; Bilitewski,
849 B., Darbra, R. M., Barceló, D., Eds.; The Handbook of Environmental Chemistry; Springer Berlin
850 Heidelberg: Berlin, Heidelberg, 2011; Vol. 18, p 290. [https://doi.org/10.1016/0143-1471\(82\)90111-8](https://doi.org/10.1016/0143-1471(82)90111-8).
- 851 (72) Robinson, T.; McMullan, G.; Marchant, R.; Nigam, P. Remediation of Dyes in Textile Effluent: A
852 Critical Review on Current Treatment Technologies with a Proposed Alternative. *Bioresour. Technol.*
853 **2001**, *77* (3), 247–255. [https://doi.org/10.1016/S0960-8524\(00\)00080-8](https://doi.org/10.1016/S0960-8524(00)00080-8).
- 854 (73) Pokharia, A.; Singh, S. Toxicological Effect of Textile Dyes and Their Metabolites: A Review. *Curr.*
855 *Trends Biotechnol. Chem. Res.* **2015**, *5* (1), 11–17.
- 856 (74) Remy, F.; Collard, F.; Gilbert, B.; Compère, P.; Eppe, G.; Lepoint, G. When Microplastic Is Not
857 Plastic: The Ingestion of Artificial Cellulose Fibers by Macrofauna Living in Seagrass
858 Macrophytodetritus. *Environ. Sci. Technol.* **2015**, *49* (18), 11158–11166.
859 <https://doi.org/10.1021/acs.est.5b02005>.
- 860 (75) Andrady, A. L. Microplastics in the Marine Environment. *Mar. Pollut. Bull.* **2011**, *62* (8), 1596–1605.
861 <https://doi.org/10.1016/j.marpolbul.2011.05.030>.
- 862 (76) Renner, G.; Schmidt, T. C.; Schram, J. Analytical Methodologies for Monitoring

- 863 Micro(Nano)Plastics: Which Are Fit for Purpose? *Curr. Opin. Environ. Sci. Heal.* **2018**, *1*, 55–61.
864 <https://doi.org/10.1016/j.coesh.2017.11.001>.
- 865 (77) Collard, F.; Gilbert, B.; Eppe, G.; Roos, L.; Compère, P.; Das, K.; Parmentier, E. Morphology of the
866 Filtration Apparatus of Three Planktivorous Fishes and Relation with Ingested Anthropogenic
867 Particles. *Mar. Pollut. Bull.* **2017**, *116* (1–2), 182–191.
868 <https://doi.org/10.1016/j.marpolbul.2016.12.067>.
- 869 (78) Löder, M. G. J.; Kuczera, M.; Mintenig, S.; Lorenz, C.; Gerdt, G. Focal Plane Array Detector-Based
870 Micro-Fourier-Transform Infrared Imaging for the Analysis of Microplastics in Environmental
871 Samples. *Environ. Chem.* **2015**, *12* (5), 563. <https://doi.org/10.1071/EN14205>.
- 872 (79) Song, Y. K.; Hong, S. H.; Jang, M.; Han, G. M.; Rani, M.; Lee, J.; Shim, W. J. A Comparison of
873 Microscopic and Spectroscopic Identification Methods for Analysis of Microplastics in
874 Environmental Samples. *Mar. Pollut. Bull.* **2015**, *93* (1–2), 202–209.
875 <https://doi.org/10.1016/j.marpolbul.2015.01.015>.
- 876 (80) GESAMP. *Sources, Fate and Effects of Microplastics in the Marine Environment: Part Two of a*
877 *Global Assessment*; Kershaw, P. J., Rochman, C. M., Eds.; Rep. Stud. GESAMP No. 93, 2016.
- 878 (81) Roch, S.; Walter, T.; Ittner, L. D.; Friedrich, C.; Brinker, A. A Systematic Study of the Microplastic
879 Burden in Freshwater Fishes of South-Western Germany - Are We Searching at the Right Scale? *Sci.*
880 *Total Environ.* **2019**, *689*, 1001–1011. <https://doi.org/10.1016/j.scitotenv.2019.06.404>.
- 881 (82) Vandermeersch, G.; Van Cauwenberghe, L.; Janssen, C. R.; Marques, A.; Granby, K.; Fait, G.;
882 Kotterman, M. J. J.; Diogène, J.; Bekaert, K.; Robbens, J.; et al. A Critical View on Microplastic
883 Quantification in Aquatic Organisms. *Environ. Res.* **2015**, *143*, 46–55.
884 <https://doi.org/10.1016/j.envres.2015.07.016>.
- 885 (83) Dümichen, E.; Barthel, A.-K.; Braun, U.; Bannick, C. G.; Brand, K.; Jekel, M.; Senz, R. Analysis of
886 Polyethylene Microplastics in Environmental Samples, Using a Thermal Decomposition Method.
887 *Water Res.* **2015**, *85*, 451–457. <https://doi.org/10.1016/j.watres.2015.09.002>.
- 888 (84) Käßler, A.; Fischer, D.; Oberbeckmann, S.; Schernewski, G.; Labrenz, M.; Eichhorn, K.-J.; Voit, B.
889 Analysis of Environmental Microplastics by Vibrational Microspectroscopy: FTIR, Raman or Both?
890 *Anal. Bioanal. Chem.* **2016**, *408* (29), 8377–8391. <https://doi.org/10.1007/s00216-016-9956-3>.
- 891 (85) Araujo, C. F.; Nolasco, M. M.; Ribeiro, A. M. P.; Ribeiro-Claro, P. J. A. Identification of

- 892 Microplastics Using Raman Spectroscopy: Latest Developments and Future Prospects. *Water Res.*
893 **2018**, *142*, 426–440. <https://doi.org/10.1016/J.WATRES.2018.05.060>.
- 894 (86) Steer, M.; Cole, M.; Thompson, R. C.; Lindeque, P. K. Microplastic Ingestion in Fish Larvae in the
895 Western English Channel. *Environ. Pollut.* **2017**, *226*, 250–259.
896 <https://doi.org/10.1016/j.envpol.2017.03.062>.
- 897 (87) Bessa, F.; Barría, P.; Neto, J. M.; Frias, J. P. G. L.; Otero, V.; Sobral, P.; Marques, J. C. Occurrence
898 of Microplastics in Commercial Fish from a Natural Estuarine Environment. *Mar. Pollut. Bull.* **2018**,
899 *128*, 575–584. <https://doi.org/10.1016/j.marpolbul.2018.01.044>.
- 900 (88) Wieczorek, A. M.; Morrison, L.; Croot, P. L.; Allcock, A. L.; MacLoughlin, E.; Savard, O.;
901 Brownlow, H.; Doyle, T. K. Frequency of Microplastics in Mesopelagic Fishes from the Northwest
902 Atlantic. *Front. Mar. Sci.* **2018**, *5*. <https://doi.org/10.3389/fmars.2018.00039>.
- 903 (89) Watts, A. J. R.; Lewis, C.; Goodhead, R. M.; Beckett, S. J.; Moger, J.; Tyler, C. R.; Galloway, T. S.
904 Uptake and Retention of Microplastics by the Shore Crab *Carcinus Maenas*. *Environ. Sci. Technol.*
905 **2014**, *48* (15), 8823–8830.
- 906 (90) Grigorakis, S.; Mason, S. A.; Drouillard, K. G. Determination of the Gut Retention of Plastic
907 Microbeads and Microfibers in Goldfish (*Carassius Auratus*). *Chemosphere* **2017**, *169*, 233–238.
908 <https://doi.org/http://dx.doi.org/10.1016/j.chemosphere.2016.11.055>.
- 909 (91) McNeish, R. E.; Kim, L. H.; Barrett, H. A.; Mason, S. A.; Kelly, J. J.; Hoellein, T. J. Microplastic in
910 Riverine Fish Is Connected to Species Traits. *Sci. Rep.* **2018**, *8* (1), 11639.
911 <https://doi.org/10.1038/s41598-018-29980-9>.
- 912 (92) Wesch, C.; Elert, A. M.; Wörner, M.; Braun, U.; Klein, R.; Paulus, M. Assuring Quality in
913 Microplastic Monitoring: About the Value of Clean-Air Devices as Essentials for Verified Data. *Sci.*
914 *Rep.* **2017**, *7* (1), 5424. <https://doi.org/10.1038/s41598-017-05838-4>.
- 915 (93) Kosuth, M.; Mason, S. A.; Wattenberg, E. V. Anthropogenic Contamination of Tap Water, Beer, and
916 Sea Salt. *PLoS One* **2018**, *13* (4), e0194970. <https://doi.org/10.1371/journal.pone.0194970>.
- 917 (94) Yang, D.; Shi, H.; Li, L.; Li, J.; Jabeen, K.; Kolandhasamy, P. Microplastic Pollution in Table Salts
918 from China. *Environ. Pollut.* **2015**, *49*, 13622–13627. <https://doi.org/10.1021/acs.est.5b03163>.
- 919 (95) Iñiguez, M. E.; Conesa, J. A.; Fullana, A. Microplastics in Spanish Table Salt. *Sci. Rep.* **2017**, *7* (1),
920 8620. <https://doi.org/10.1038/s41598-017-09128-x>.

- 921 (96) Campbell, S. H.; Williamson, P. R.; Hall, B. D. Microplastics in the Gastrointestinal Tracts of Fish
922 and the Water from an Urban Prairie Creek. *FACETS* **2017**, *2* (1), 395–409.
923 <https://doi.org/10.1139/facets-2017-0008>.
- 924 (97) Possatto, F. E.; Barletta, M.; Costa, M. F.; do Sul, J. A. I.; Dantas, D. V. Plastic Debris Ingestion by
925 Marine Catfish: An Unexpected Fisheries Impact. *Mar. Pollut. Bull.* **2011**, *62* (5), 1098–1102.
926 <https://doi.org/10.1016/j.marpolbul.2011.01.036>.
- 927 (98) Sanchez, W.; Bender, C.; Porcher, J.-M. Wild Gudgeons (*Gobio Gobio*) from French Rivers Are
928 Contaminated by Microplastics: Preliminary Study and First Evidence. *Environ. Res.* **2014**, *128*, 98–
929 100. <https://doi.org/10.1016/j.envres.2013.11.004>.
- 930 (99) Biginagwa, F. J.; Mayoma, B. S.; Shashoua, Y.; Syberg, K.; Khan, F. R. First Evidence of
931 Microplastics in the African Great Lakes: Recovery from Lake Victoria Nile Perch and Nile Tilapia.
932 *J. Great Lakes Res.* **2016**, *42* (1). <https://doi.org/10.1016/j.jglr.2015.10.012>.
- 933 (100) Faure, F.; Saini, C.; Potter, G.; Galgani, F.; de Alencastro, L. F.; Haggmann, P. An Evaluation of
934 Surface Micro- and Mesoplastic Pollution in Pelagic Ecosystems of the Western Mediterranean Sea.
935 *Environ. Sci. Pollut. Res. Int.* **2015**. <https://doi.org/10.1007/s11356-015-4453-3>.
- 936 (101) Phillips, M. B.; Bonner, T. H. Occurrence and Amount of Microplastic Ingested by Fishes in
937 Watersheds of the Gulf of Mexico. *Mar. Pollut. Bull.* **2015**.
938 <https://doi.org/10.1016/j.marpolbul.2015.08.041>.
- 939 (102) Ferreira, G. V. B.; Barletta, M.; Lima, A. R. A.; Dantas, D. V.; Justino, A. K. S.; Costa, M. F. Plastic
940 Debris Contamination in the Life Cycle of Acoupa Weakfish (*Cynoscion Acoupa*) in a Tropical
941 Estuary. *ICES J. Mar. Sci. J. du Cons.* **2016**, *73* (10), 2695–2707.
942 <https://doi.org/10.1093/icesjms/fsw108>.
- 943 (103) Peters, C. A.; Bratton, S. P. Urbanization Is a Major Influence on Microplastic Ingestion by Sunfish
944 in the Brazos River Basin, Central Texas, USA. *Environ. Pollut.* **2016**, *210*, 380–387.
945 <https://doi.org/10.1016/j.envpol.2016.01.018>.
- 946 (104) McGoran, A. R.; Clark, P. F.; Morrith, D. Presence of Microplastic in the Digestive Tracts of
947 European Flounder, *Platichthys Flesus*, and European Smelt, *Osmerus Eperlanus*, from the River
948 Thames. *Environ. Pollut.* **2017**, *220*, 744–751. <https://doi.org/10.1016/j.envpol.2016.09.078>.
- 949 (105) Silva-Cavalcanti, J. S.; Silva, J. D. B.; França, E. J. de; Araújo, M. C. B. de; Gusmão, F.

- 950 Microplastics Ingestion by a Common Tropical Freshwater Fishing Resource. *Environ. Pollut.* **2017**,
951 *221*, 218–226. <https://doi.org/10.1016/j.envpol.2016.11.068>.
- 952 (106) Abbasi, S.; Soltani, N.; Keshavarzi, B.; Moore, F.; Turner, A.; Hassanaghahi, M. Microplastics in
953 Different Tissues of Fish and Prawn from the Musa Estuary, Persian Gulf. *Chemosphere* **2018**, *205*,
954 80–87. <https://doi.org/10.1016/j.chemosphere.2018.04.076>.
- 955 (107) Xiong, X.; Zhang, K.; Chen, X.; Shi, H.; Luo, Z.; Wu, C. Sources and Distribution of Microplastics in
956 China's Largest Inland Lake – Qinghai Lake. *Environ. Pollut.* **2018**, *235*, 899–906.
957 <https://doi.org/10.1016/j.envpol.2017.12.081>.
- 958 (108) Andrade, M. C.; Winemiller, K. O.; Barbosa, P. S.; Fortunati, A.; Chelazzi, D.; Cincinelli, A.;
959 Giarrizzo, T. First Account of Plastic Pollution Impacting Freshwater Fishes in the Amazon:
960 Ingestion of Plastic Debris by Piranhas and Other Serrasalminids with Diverse Feeding Habits.
961 *Environ. Pollut.* **2019**, *244*, 766–773. <https://doi.org/10.1016/j.envpol.2018.10.088>.
- 962 (109) Sloommaekers, B.; Catarci Carteny, C.; Belpaire, C.; Saverwyns, S.; Fremout, W.; Blust, R.; Bervoets,
963 L. Microplastic Contamination in Gudgeons (*Gobio Gobio*) from Flemish Rivers (Belgium). *Environ.*
964 *Pollut.* **2019**, *244*, 675–684. <https://doi.org/10.1016/j.envpol.2018.09.136>.
- 965 (110) Yuan, W.; Liu, X.; Wang, W.; Di, M.; Wang, J. Microplastic Abundance, Distribution and
966 Composition in Water, Sediments, and Wild Fish from Poyang Lake, China. *Ecotoxicol. Environ. Saf.*
967 **2019**, *170*, 180–187. <https://doi.org/10.1016/j.ecoenv.2018.11.126>.
- 968 (111) Su, L.; Deng, H.; Li, B.; Chen, Q.; Pettigrove, V.; Wu, C.; Shi, H. The Occurrence of Microplastic in
969 Specific Organs in Commercially Caught Fishes from Coast and Estuary Area of East China. *J.*
970 *Hazard. Mater.* **2019**, *365*, 716–724. <https://doi.org/10.1016/j.jhazmat.2018.11.024>.
- 971 (112) Ferreira, G. V. B.; Barletta, M.; Lima, A. R. A. Use of Estuarine Resources by Top Predator Fishes.
972 How Do Ecological Patterns Affect Rates of Contamination by Microplastics? *Sci. Total Environ.*
973 **2019**, *655*, 292–304. <https://doi.org/10.1016/j.scitotenv.2018.11.229>.
- 974 (113) Arias, A. H.; Ronda, A. C.; Oliva, A. L.; Marcovecchio, J. E. Evidence of Microplastic Ingestion by
975 Fish from the Bahía Blanca Estuary in Argentina, South America. *Bull. Environ. Contam. Toxicol.*
976 **2019**. <https://doi.org/10.1007/s00128-019-02604-2>.
- 977 (114) Su, L.; Nan, B.; Hassell, K. L.; Craig, N. J.; Pettigrove, V. Microplastics Biomonitoring in Australian
978 Urban Wetlands Using a Common Noxious Fish (*Gambusia Holbrooki*). *Chemosphere* **2019**, *228*,

- 979 65–74. <https://doi.org/10.1016/j.chemosphere.2019.04.114>.
- 980 (115) Hidalgo-Ruz, V.; Gutow, L.; Thompson, R. C.; Thiel, M. Microplastics in the Marine Environment:
981 A Review of the Methods Used for Identification and Quantification. *Environ. Sci. Technol.* **2012**, *46*
982 (6), 3060–3075.
- 983 (116) Zobkov, M. B.; Esiukova, E. E. Microplastics in a Marine Environment: Review of Methods for
984 Sampling, Processing, and Analyzing Microplastics in Water, Bottom Sediments, and Coastal
985 Deposits. *Oceanology* **2018**, *58* (1), 137–143. <https://doi.org/10.1134/S0001437017060169>.
- 986 (117) Rodrigues, M. O.; Abrantes, N.; Gonçalves, F. J. M.; Nogueira, H.; Marques, J. C.; Gonçalves, A. M.
987 M. Spatial and Temporal Distribution of Microplastics in Water and Sediments of a Freshwater
988 System (Antuã River, Portugal). *Sci. Total Environ.* **2018**, *633*, 1549–1559.
989 <https://doi.org/10.1016/j.scitotenv.2018.03.233>.
- 990 (118) Lin, L.; Zuo, L.-Z.; Peng, J.-P.; Cai, L.-Q.; Fok, L.; Yan, Y.; Li, H.-X.; Xu, X.-R. Occurrence and
991 Distribution of Microplastics in an Urban River: A Case Study in the Pearl River along Guangzhou
992 City, China. *Sci. Total Environ.* **2018**, *644*, 375–381. <https://doi.org/10.1016/j.scitotenv.2018.06.327>.
- 993 (119) Su, L.; Xue, Y.; Li, L.; Yang, D.; Kolandhasamy, P.; Li, D.; Shi, H. Microplastics in Taihu Lake,
994 China. *Environ. Pollut.* **2016**, *216*, 711–719. <https://doi.org/10.1016/j.envpol.2016.06.036>.
- 995 (120) Ballent, A.; Corcoran, P. L.; Madden, O.; Helm, P. A.; Longstaffe, F. J. Sources and Sinks of
996 Microplastics in Canadian Lake Ontario Nearshore, Tributary and Beach Sediments. *Mar. Pollut.*
997 *Bull.* **2016**, *110* (1), 383–395. <https://doi.org/10.1016/j.marpolbul.2016.06.037>.
- 998 (121) McCormick, A.; Hoellein, T. J.; Mason, S. A.; Schlupe, J.; Kelly, J. J. Microplastic Is an Abundant
999 and Distinct Microbial Habitat in an Urban River. *Environ. Sci. Technol.* **2014**, *48* (20), 11863–
1000 11871. <https://doi.org/10.1021/es503610r>.
- 1001 (122) Claessens, M.; Meester, S. D.; Landuyt, L. V.; Clerck, K. D.; Janssen, C. R. Occurrence and
1002 Distribution of Microplastics in Marine Sediments along the Belgian Coast. *Mar. Pollut. Bull.* **2011**,
1003 *62* (10), 2199–2204.
- 1004 (123) Vianello, A.; Boldrin, A.; Guerriero, P.; Moschino, V.; Rella, R.; Sturaro, A.; Da Ros, L. Microplastic
1005 Particles in Sediments of Lagoon of Venice, Italy: First Observations on Occurrence, Spatial Patterns
1006 and Identification. *Estuar. Coast. Shelf Sci.* **2013**, *130*, 54–61.
1007 <https://doi.org/10.1016/j.ecss.2013.03.022>.

- 1008 (124) Kim, S. W.; Chae, Y.; Kim, D.; An, Y.-J. Zebrafish Can Recognize Microplastics as Inedible
1009 Materials: Quantitative Evidence of Ingestion Behavior. *Sci. Total Environ.* **2019**, *649*, 156–162.
1010 <https://doi.org/10.1016/j.scitotenv.2018.08.310>.
- 1011 (125) Vroom, R. J. E.; Koelmans, A. A.; Besseling, E.; Halsband, C. Aging of Microplastics Promotes
1012 Their Ingestion by Marine Zooplankton. *Environ. Pollut.* **2017**, *231, Part*, 987–996.
1013 <https://doi.org/https://doi.org/10.1016/j.envpol.2017.08.088>.
- 1014 (126) Savoca, M. S.; Wohlfeil, M. E.; Ebeler, S. E.; Nevitt, G. A. Marine Plastic Debris Emits a Keystone
1015 Infochemical for Olfactory Foraging Seabirds. *Sci. Adv.* **2016**, *2* (11).
- 1016 (127) Savoca, M. S.; Tyson, C. W.; McGill, M.; Slager, C. J. Odours from Marine Plastic Debris Induce
1017 Food Search Behaviours in a Forage Fish. *Proc. R. Soc. B Biol. Sci.* **2017**, *284* (1860), 20171000.
1018 <https://doi.org/10.1098/rspb.2017.1000>.
- 1019 (128) Cheung, P. K.; Cheung, L. T. O.; Fok, L. Seasonal Variation in the Abundance of Marine Plastic
1020 Debris in the Estuary of a Subtropical Macro-Scale Drainage Basin in South China. *Sci. Total*
1021 *Environ.* **2016**, *562*, 658–665. <https://doi.org/10.1016/j.scitotenv.2016.04.048>.
- 1022 (129) Goldberg, E. D. Plasticizing the Seafloor: An Overview. *Environ. Technol.* **1997**, *18* (2), 195–201.
1023 <https://doi.org/10.1080/09593331808616527>.
- 1024 (130) Lusher, A. L.; McHugh, M.; Thompson, R. C. Occurrence of Microplastics in the Gastrointestinal
1025 Tract of Pelagic and Demersal Fish from the English Channel. *Mar. Pollut. Bull.* **2013**, *67* (1–2), 94–
1026 99.
- 1027 (131) Chagnon, C.; Thiel, M.; Antunes, J.; Ferreira, J. L.; Sobral, P.; Ory, N. C. Plastic Ingestion and
1028 Trophic Transfer between Easter Island Flying Fish (*Cheilopogon Rapanouiensis*) and Yellowfin
1029 Tuna (*Thunnus Albacares*) from Rapa Nui (Easter Island). *Environ. Pollut.* **2018**, *243*, 127–133.
1030 <https://doi.org/10.1016/j.envpol.2018.08.042>.
- 1031 (132) Bravo Rebolledo, E. L.; Van Franeker, J. A.; Jansen, O. E.; Brasseur, S. M. J. M. Plastic Ingestion by
1032 Harbour Seals (*Phoca Vitulina*) in The Netherlands. *Mar. Pollut. Bull.* **2013**, *67* (1–2), 200–202.
1033 <https://doi.org/10.1016/j.marpolbul.2012.11.035>.
- 1034 (133) Donohue, M. J.; Masura, J.; Gelatt, T.; Ream, R.; Baker, J. D.; Faulhaber, K.; Lerner, D. T.
1035 Evaluating Exposure of Northern Fur Seals, *Callorhinus Ursinus*, to Microplastic Pollution through
1036 Fecal Analysis. *Mar. Pollut. Bull.* **2019**, *138*, 213–221.

- 1037 <https://doi.org/10.1016/j.marpolbul.2018.11.036>.
- 1038 (134) Clements, K. D.; Choat, J. H. Influence of Season, Ontogeny and Tide on the Diet of the Temperate
1039 Marine Herbivorous Fish *Odax Pullus* (Odacidae). *Mar. Biol.* **1993**, *117* (2), 213–220.
1040 <https://doi.org/10.1007/BF00345665>.
- 1041 (135) King, A. J. Ontogenetic Dietary Shifts of Fishes in an Australian Floodplain River. *Mar. Freshw. Res.*
1042 **2005**, *56* (2), 215. <https://doi.org/10.1071/MF04117>.
- 1043 (136) Demain, D. K.; Gallego, A.; Jaworski, A.; Priede, I. G.; Jones, E. G. Diet and Feeding Niches of
1044 Juvenile *Gadus Morhua*, *Melanogrammus Aeglefinus* and *Merlangius Merlangus* during the
1045 Settlement Transition in the Northern North Sea. *J. Fish Biol.* **2011**, *79* (1), 89–111.
1046 <https://doi.org/10.1111/j.1095-8649.2011.02997.x>.
- 1047 (137) Nunn, A. D.; Tewson, L. H.; Cowx, I. G. The Foraging Ecology of Larval and Juvenile Fishes. *Rev.*
1048 *Fish Biol. Fish.* **2012**, *22* (2), 377–408. <https://doi.org/10.1007/s11160-011-9240-8>.
- 1049 (138) Anastasopoulou, A.; Mytilineou, C.; Smith, C. J.; Papadopoulou, K. N. Plastic Debris Ingested by
1050 Deep-Water Fish of the Ionian Sea (Eastern Mediterranean). *Deep. Res. Part I Oceanogr. Res. Pap.*
1051 **2013**, *74*, 11–13.
- 1052 (139) Lazzaro, X. A Review of Planktivorous Fishes: Their Evolution, Feeding Behaviours, Selectivities,
1053 and Impacts. *Hydrobiologia* **1987**, *146* (2), 97–167. <https://doi.org/10.1007/BF00008764>.
- 1054 (140) Ory, N. C.; Gallardo, C.; Lenz, M.; Thiel, M. Capture, Swallowing, and Egestion of Microplastics by
1055 a Planktivorous Juvenile Fish. *Environ. Pollut.* **2018**, *240*, 566–573.
1056 <https://doi.org/10.1016/J.ENVPOL.2018.04.093>.
- 1057 (141) Carson, H. S. The Incidence of Plastic Ingestion by Fishes: From the Prey's Perspective. *Mar. Pollut.*
1058 *Bull.* **2013**.
- 1059 (142) Lima, A. R. A.; Costa, M. F.; Barletta, M. Distribution Patterns of Microplastics within the Plankton
1060 of a Tropical Estuary. *Environ. Res.* **2014**, *132*, 146–155.
1061 <https://doi.org/10.1016/j.envres.2014.03.031>.
- 1062 (143) van der Oost, R.; Beyer, J.; Vermeulen, N. P. . Fish Bioaccumulation and Biomarkers in
1063 Environmental Risk Assessment: A Review. *Environ. Toxicol. Pharmacol.* **2003**, *13* (2), 57–149.
1064 [https://doi.org/10.1016/S1382-6689\(02\)00126-6](https://doi.org/10.1016/S1382-6689(02)00126-6).
- 1065 (144) Browne, M. A.; Crump, P.; Niven, S. J.; Teuten, E.; Tonkin, A.; Galloway, T.; Thompson, R.

- 1066 Accumulation of Microplastic on Shorelines Worldwide: Sources and Sinks. *Environ. Sci. Technol.*
1067 **2011**, *45* (21), 9175–9179. <https://doi.org/10.1021/es201811s>.
- 1068 (145) Salvador Cesa, F.; Turra, A.; Baruque-Ramos, J. Synthetic Fibers as Microplastics in the Marine
1069 Environment: A Review from Textile Perspective with a Focus on Domestic Washings. *Sci. Total*
1070 *Environ.* **2017**, *598*, 1116–1129. <https://doi.org/10.1016/j.scitotenv.2017.04.172>.
- 1071 (146) Kase, R.; Korkaric, M.; Werner, I.; Ågerstrand, M. Criteria for Reporting and Evaluating Ecotoxicity
1072 Data (CRED): Comparison and Perception of the Klimisch and CRED Methods for Evaluating
1073 Reliability and Relevance of Ecotoxicity Studies. *Environ. Sci. Eur.* **2016**, *28* (1), 7.
1074 <https://doi.org/10.1186/s12302-016-0073-x>.
- 1075 (147) Collard, F.; Gilbert, B.; Compère, P.; Eppe, G.; Das, K.; Jauniaux, T.; Parmentier, E. Microplastics in
1076 Livers of European Anchovies (*Engraulis Encrasicolus*, L.). *Environ. Pollut.* **2017**, *229*, 1000–1005.
1077 <https://doi.org/10.1016/j.envpol.2017.07.089>.
- 1078 (148) Ribeiro, F.; Garcia, A. R.; Pereira, B. P.; Fonseca, M.; Mestre, N. C.; Fonseca, T. G.; Ilharco, L. M.;
1079 Bebianno, M. J. Microplastics Effects in Scrobicularia Plana. *Mar. Pollut. Bull.* **2017**, *122* (1–2),
1080 379–391. <https://doi.org/10.1016/j.marpolbul.2017.06.078>.
- 1081 (149) Khan, M. B.; Prezant, R. S. Microplastic Abundances in a Mussel Bed and Ingestion by the Ribbed
1082 Marsh Mussel *Geukensia Demissa*. *Mar. Pollut. Bull.* **2018**, *130*, 67–75.
1083 <https://doi.org/10.1016/j.marpolbul.2018.03.012>.
- 1084 (150) Rochman, C. M.; Browne, M. A.; Underwood, A. J.; van Franeker, J. A.; Thompson, R. C.; Amaral-
1085 Zettler, L. A. The Ecological Impacts of Marine Debris: Unraveling the Demonstrated Evidence from
1086 What Is Perceived. *Ecology* **2016**, *97* (2), 302–312. <https://doi.org/10.1890/14-2070.1>.
- 1087 (151) Mazurais, D.; Ernande, B.; Quazuguel, P.; Severe, A.; Huelvan, C.; Madec, L.; Mouchel, O.;
1088 Soudant, P.; Robbens, J.; Huvet, A.; et al. Evaluation of the Impact of Polyethylene Microbeads
1089 Ingestion in European Sea Bass (*Dicentrarchus Labrax*) Larvae. *Mar. Environ. Res.* **2015**, *112*, 78–
1090 85. <https://doi.org/10.1016/j.marenvres.2015.09.009>.
- 1091 (152) de Sá, L. C.; Oliveira, M.; Ribeiro, F.; Rocha, T. L.; Futter, M. N. Studies of the Effects of
1092 Microplastics on Aquatic Organisms: What Do We Know and Where Should We Focus Our Efforts
1093 in the Future? *Sci. Total Environ.* **2018**, *645*, 1029–1039.
1094 <https://doi.org/10.1016/j.scitotenv.2018.07.207>.

- 1095 (153) Crain, D. A.; Eriksen, M.; Iguchi, T.; Jobling, S.; Laufer, H.; LeBlanc, G. A.; Guillette, L. J. An
1096 Ecological Assessment of Bisphenol-A: Evidence from Comparative Biology. *Reprod. Toxicol.* **2007**,
1097 *24* (2), 225–239. <https://doi.org/10.1016/j.reprotox.2007.05.008>.
- 1098 (154) Oehlmann, J.; Schulte-Oehlmann, U.; Kloas, W.; Jagnytsch, O.; Lutz, I.; Kusk, K. O.; Wollenberger,
1099 L.; Santos, E. M.; Paull, G. C.; Van Look, K. J. W.; et al. A Critical Analysis of the Biological
1100 Impacts of Plasticizers on Wildlife. *Philos. Trans. R. Soc. Lond. B. Biol. Sci.* **2009**, *364* (1526), 2047–
1101 2062. <https://doi.org/10.1098/rstb.2008.0242>.
- 1102 (155) Johnson, L. L.; Anulacion, B. F.; Arkoosh, M. R.; Burrows, D. G.; da Silva, D. A. M.; Dietrich, J. P.;
1103 Myers, M. S.; Spromberg, J.; Ylitalo, G. M. Effects of Legacy Persistent Organic Pollutants (POPs)
1104 in Fish—Current and Future Challenges. In *Organic Chemical Toxicology of fishes*; Tierney, K. B.,
1105 Farrell, A. P., Brauner, C. J., Eds.; Elsevier, 2013; pp 53–140. [https://doi.org/10.1016/B978-0-12-](https://doi.org/10.1016/B978-0-12-398254-4.00002-9)
1106 [398254-4.00002-9](https://doi.org/10.1016/B978-0-12-398254-4.00002-9).
- 1107