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1     **Kinetics of methane hydrate formation and dissociation in**  
2                                     **sand sediment**

3

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15 **Abstract**

16 Methane hydrate is being considered as a potential future energy source but also a  
17 considerable geo-hazard. In this study, methane hydrate bearing sand sediment was  
18 firstly created by pressurizing methane gas into already chilled moistened packed sand  
19 specimen (excess gas method). The excess gas was then replaced by water at high  
20 pressure. Afterward, a heating/cooling cycle was applied under undrained conditions  
21 in order to completely dissociate gas hydrates and then recreate them inside the  
22 specimen. Finally, the pore pressure was reduced to zero to dissociate the gas  
23 hydrates. The whole process was performed in a magnetic resonance imaging (MRI)  
24 system allowing the determination of water and/or gas and hydrate quantity (and  
25 spatial distribution) at various times. The MRI signal was finally analyzed to interpret  
26 various processes in sand sediment: initial hydrate formation, heating-induced hydrate  
27 dissociation, cooling-induced hydrate re-formation, and depressurizing-induced  
28 hydrate dissociation.

29

30 **Keyword:** gas hydrate bearing sand sediment, dissociation/formation, kinetics,  
31 magnetic resonance imaging.

32

## 33 **1. Introduction**

34 Natural gas hydrates (primarily methane hydrates forming naturally at high pressures  
35 and low temperatures) are nowadays being considered as an alternative energy  
36 source [1]. Among the existing methods of gas recovery from hydrates,  
37 depressurization method is being considered as the most economically promising  
38 method [2]. This method, conducted by lowering the pressure in overlying sediments,  
39 may be hampered by the formation of ice and/or the reformation of gas hydrates (GH)  
40 because of the endothermic cooling nature of GH. Fundamental understandings of  
41 hydrate dissociation kinetics models are essential to predict hydrate reservoir  
42 dissociation process in the objective of selecting appropriate hydrate bearing zones  
43 and estimating gas production behavior before execution of any field tests. Some  
44 kinetics models were developed to simulate the production process based on  
45 heat/mass transfer and/or intrinsic kinetics of hydrate decomposition and/or gas-water  
46 two-phase flow [3,4]. Different assumptions were used, their applicability to reservoir  
47 level studies is also limited case by case. Various GH reservoir simulators  
48 (computational tools taking into account the complex highly-coupled transport  
49 equations, the reaction kinetics, the phase transition and the physical/chemical  
50 properties of hydrate bearing sediment) are being developed [5,6]. The accuracy of  
51 reservoir model needs, however, to be improved and the availability of long-term field  
52 production test data is so important. Two successful offshore field tests (together with  
53 two onshore field tests in Mount Elbert – Alaska and Malik in 2007 [7]), up to now, used  
54 the depressurization method. The first offshore methane hydrate production test was  
55 conducted by Japan Oil, Gas, and Metals National Corporation (JOGMEC) in the  
56 eastern Nankai Trough. Approximately 120,000 m<sup>3</sup> of methane gas (20,000 m<sup>3</sup>/day)  
57 was produced by lowering pressure from 13.5 MPa to 4.5 MPa. The production was

58 interrupted due to an unexpected increase in sand production [8]. Recently in 2017,  
59 the China Geological Survey extracted 120,000 m<sup>3</sup> of gas from natural gas hydrate  
60 deposits in the Shenhu area with a methane concentration of 99.5 percent (Xin Z.  
61 China wraps up combustible ice mining trial, setting world records. Xinhua.  
62 Guangzhou, 09 July 2017). As production costs are still high, an economically feasible  
63 way to exploit gas hydrate on a large scale should be found to commercialize the  
64 production of the natural gas hydrate. As it is challenging to get intact cores of methane  
65 hydrate-bearing sediments, most the experimental works concern laboratory tests on  
66 synthetic specimens to investigate hydrate dissociation rate, pressure-temperature  
67 evolution [9–13]. The experimental reactor scale is a crucial factor; a larger reactor  
68 better mimics actual field conditions but it would be more difficult to ensure the  
69 homogeneity of synthesized specimens [14].

70

71 In the objective of creating synthetic methane hydrates in sandy sediments, some  
72 methods were proposed and well-studied as: dissolved gas [15], partial water  
73 saturation [16], excess water [17] or ice-seeding [18]. Among them, dissolved gas and  
74 water excess method are supposed to form non-cementing hydrate habit in sandy  
75 sediments. However, dissolved gas is time-consuming method especially at high  
76 hydrate saturation because of low solubility of methane gas in water [15]. In addition,  
77 methane hydrate is observed form heterogeneously inside the sample by using water  
78 excess method by mean of X-Ray Micro-Tomography [19] and measures of pressure  
79 at different positions in the sample [20]. Recently, Choi et al. [21] proposed an efficient  
80 and consistent method by combining the partial water saturation, saline water injection  
81 at restricted conditions and a temperature cycle. However,  $V_P$  after the heating process

82 is quite high while sample is not saturated. The hydrate dissociation is perhaps not  
83 finished before the hydrate reformation.

84

85 Besides elastic wave velocity measurement [18,22–24] and synchrotron X-ray  
86 computed tomographic microscopy [25,26], which are used to study the kinetics and  
87 mechanisms of hydrate formation and dissociation, <sup>1</sup>H Nuclear magnetic resonance  
88 spectroscopy (NMR), in particular Magnetic Resonance Imaging (MRI) (at  
89 macroscopic scale of NMR) is a well-suited mean to quantitatively/qualitatively follow  
90 these kinetics. In most MRI studied cases, glass beads were used to simulate the  
91 porous media to investigate the tetrahydrofuran (THF), carbon dioxide (CO<sub>2</sub>) hydrate  
92 formation and dissociation [27,28]. Methane hydrate formation (by dissolved gas or  
93 partially water saturation methods) was observed via Mean Intensity (*MI*) evolution and  
94 2D images [29,30]. Effects of different sizes of glass beads on hydrate growth stage  
95 were investigated. In general, hydrate growth rate increases when the size of the  
96 porous media decreases. In addition, three growth stages of methane hydrates formed  
97 after the partially water saturation method were observed: the initial growth, the rapid  
98 growth and the steady stage. In sandstone media, methane hydrate formation and  
99 spontaneous conversion of methane to CO<sub>2</sub> hydrate were studied by mean of *MI* and  
100 3D images [31,32]. As time is needed to take 3D images, spatial distribution of  
101 specimen during the hydrate formation was not measured regularly. The formation of  
102 methane hydrates in unconsolidated bed of silica with different size ranges was  
103 investigated by following the *MI* evolution, measured on vertical and horizontal slices  
104 [33]. The formation was observed non-uniform and occurred at different times and  
105 different positions. In addition, by using different water saturations, hydrate formation  
106 was found to be faster for lower water content. The methane hydrate formation,

107 dissociation and reformation in partially water saturated Ottawa sand at different water  
108 saturations were studied by combining measurements of  $M/I$  and elastic velocities [34].  
109  $M/I$  profiles along the specimen axis after these three procedures (hydrate formation,  
110 dissociation and reformation) show an almost homogenous distribution of GH. The fact  
111 of consolidating unsaturated sand makes water distributed more homogeneously before  
112 the gas hydrate formation. The effect of depressurizing range and rate on methane  
113 hydrate dissociation, in particular, the hydrate reformation and ice generation due to  
114 fast depressurizing rate were observed [35,36].

115

116 In this study, methane hydrate formation based on the method proposed by Choi et al.  
117 [21], but modified to improve the heating-cooling process and to adapt better with  
118 existing facilities is investigated in the objective of following the specimen homogeneity  
119 during the whole GH formation phase. In addition, GH dissociation after  
120 depressurization method is observed. Methodological efforts were put in getting fast  
121 enough measurements to follow the kinetics of GH formation/dissociation during  
122 transitory steps.

## 123 **2. Experimental method**

### 124 **2.1. Materials**

125 The soil used in this study is Fontainebleau silica sand (NE34). It consists of poor-  
126 graded sub-rounded grains having diameter ranging from 100 to 300 microns (see the  
127 grain size distribution curve in Figure 1 obtained by laser diffraction analysis). The  
128 physical characteristics of this material are detailed in Table 1. Tap water and methane  
129 gas with 99.995% of purity were used in the tests.

## 130        **2.2.        Experimental setup**

131        The schematic views of experimental setup are presented in Figure 2. The sand  
132        specimen (1), 38 mm in diameter and 76 mm in height, is covered with a neoprene  
133        membrane (2). The confining pressure is applied to the specimen by a  
134        volume/pressure controller (7) using a perfluorinated oil (Galden®) as confining fluid  
135        (3), chosen due to its low signal intensity in MRI measurements. Methane gas is  
136        injected via the bottom inlet (5) by a pressure controller connected to a gas flowmeter  
137        (10). The top inlet (6) is closed in this study. A second volume/pressure controller (12)  
138        is used to control the water pore pressure. The specimen temperature is controlled by  
139        circulating a perfluorinated oil (Galden®), which is connected to a cryostat (8), around  
140        the cell (4). The cell is installed in a nuclear magnetic resonance imaging system (13)  
141        for observations.

142

143        Proton ( $^1\text{H}$ ) NMR/MRI measurements are performed at a Bruker 24/80 DBX  
144        spectrometer operating at 0.5T (21MHz proton frequency) equipped with:

145        -        A birdcage RF coil 20cm in diameter and height where the whole pressure cell  
146        can fit

147        -        A BGA-26 gradient system delivering a maximum gradient strength of 50mT/m  
148        with a rising time of 500 $\mu\text{s}$ .

149

150        Measurement protocols used in this work rely on well-established methodology. They  
151        consist of:

152        -        A pulse acquisition sequence, where the overall NMR signal owing to hydrogen  
153        is measured after a dead time of 40 $\mu\text{s}$  following the exciting RF pulse. This  
154        signal is referred to as 'FID INTENSITY' signal hereafter.



155 - A 1D profile imaging based on spin-echo acquisition with a read-out gradient  
156 orientated in the vertical direction and an echo time of 4.2ms, which provides  
157 profile measurements with 200 pixels covering a field of view of 20cm, being  
158 large enough to avoid any image aliasing owing to some parts of the external  
159 set-up to the observation zone. It provides a space-resolved view of the  
160 contribution

161 In both kinds of measurement, the signal intensity is expected to be proportional to the  
162 amount of hydrogen atoms owing to either liquid (water) or gas (methane) phases.  
163 Note that due to the Curie-law for spin polarization, the signal intensity is also inversely  
164 proportional to the absolute temperature in °K of the sample. The dead-time and the  
165 echo time are regarded as short enough to neglect bias owing to spin-spin relaxation.  
166 On the contrary, the gas-hydrate phase, and ice phase are negligible due to its short  
167 spin-spin relaxation time. Let us emphasize that FID intensities do not correspond  
168 directly to profile intensities, since the integration relationship between them depends  
169 on other parameters such as the sample size and the fluid amount out of the sample.  
170 Related data are then presented on independent scales.

171

172 If any, the related data processing relied on home-made routines under Scilab.

### 173 **2.3. Test procedure**

174 Methane hydrate bearing sediment (MHBS) specimens were prepared by the following  
175 procedure:

176 - Step 1 - Compaction: A determined quantity of moist sand (having known  
177 moisture content) is tamped in layers to obtain a specimen with a void ratio of  
178 0.63 inside the neoprene membrane before the assembly of the experimental  
179 setup as shown in Figure 2.

- 180 - Step 2 - Consolidation: The confining pressure is increased to 25 MPa then  
181 decreased to 10 MPa).
- 182 - Step 3 – Hydrate creation: The temperature of the cell is decreased to 2 °C.  
183 Vacuum is then applied to eliminate pore air in the specimen prior to the injection  
184 of methane gas at 7 MPa of pressure.
- 185 - Step 4 – Water saturation: The valve  $V_2$  is opened to atmosphere during a short  
186 period to let all the excess methane gas (initially under a pressure of 7 MPa)  
187 escape from the specimen (pore pressure decreases to zero), this valve is  
188 closed after that. Expecting that gas hydrates do not dissociate during this short  
189 period. The valve  $V_1$  is then immediately opened, the bottom inlet is connected  
190 to the volume/pressure controller (12) to inject tap water (at ambient  
191 temperature) fixed at 7 MPa of pressure to the sample. This procedure is used  
192 to replace the excess gas in the specimen by water while minimizing the  
193 disturbance of methane hydrates that already exist inside the specimen. Gas  
194 hydrate pressure – temperature conditions are the same as that at the end of  
195 step 2. Skempton's coefficient is measured at the end of this step to make sure  
196 that the sample is fully saturated.
- 197 - Step 5 – Heating-induced hydrate dissociation: From this step, the confining  
198 pressure is imposed to be always 3 MPa higher than the pore pressure. The  
199 pore pressure is first decreased from 7 MPa to 4 MPa. The drainage valve ( $V_1$ )  
200 ( $V_2$  is always closed) is then closed and the temperature of the cell is increased  
201 up to higher than 25 °C. This aims at heating the specimen under undrained  
202 condition to dissociate the existing gas hydrate progressively. Note that the pore  
203 pressure at the end of this step is higher for higher initial hydrate saturation. In  
204 addition, the capacity of pore pressure sensor in this study is limited at 19MPa.

205 The pore pressure is increased to 19 MPa at the end of this step so that this  
206 test procedure can be always feasible regardless of the hydrate saturation.

207 - Step 6 – Cooling-induced hydrate re-formation: The cell temperature is  
208 decreased to 2 °C while the pore pressure is maintained constant at 19 MPa by  
209 injecting water from the volume/pressure controller (12) into the sample. In the  
210 objectives of reforming gas hydrates faster, this provides also the same final  
211 pressure - temperature conditions for gas hydrates in the end for all tests.

212 - Step 7 – Depressurization-induced hydrate dissociation: The confining pressure  
213 is maintained at 22 MPa while the valve  $V_2$  is opened to decrease the pore  
214 pressure. The volume of methane dissociated from the specimen is measured  
215 by the system (9) composed also a gas/water separator and a gas collection  
216 system.

217 The steps 3 – 7 are performed in the MRI system and the data are logged automatically  
218 during these steps.

#### 219 **2.4. Calibration tests**

220 Calibration tests were performed at 2°C on the compacted specimen of the first test,  
221 the density of which was also very similar to that of test 2, saturated with pure phases  
222 of various fluids: (a) vacuum; (b) methane gas at 7 MPa of pressure; (c) water at 7  
223 MPa of pressure; (d) and water at 19 MPa of pressure. In Figure 3, *FID INTENSITY*  
224 obtained for the whole system in each case is plotted. The values corresponding to  
225 methane gas at 7 MPa of pressure, water at 7 MPa of pressure, and water at 19 MPa  
226 of pressure were then calculated by subtracting that corresponding to the system  
227 containing vacuum, in order to remove the spurious signal owing to the pressure cell  
228 and the imperfectly perfluorinated oil. In the working conditions of the present study,  
229 and as far as the temperature is not modified, the corrected signal is directly

230 proportional to the total amount of hydrogen atoms contained in the fluid molecules.  
231 The corrected values of *FID INTENSITY* are also plotted in the Figure 3. Note that the  
232 signal for pure methane is significantly smaller than that for water due to the different  
233 density and chemical composition. In the subsequent sections, the corrected values of  
234 *FID INTENSITY*, i.e. *FID INTENSITY* measured minus *FID INTENSITY* obtained from  
235 the case (a), are shown.

## 236 **2.5. Test program**

237 Two tests were performed in this study with the same procedure and the same  
238 parameters to ensure the repeatability of the results. The water saturation obtained  
239 after compaction equals to 25% (corresponding to a moisture content of 6%).

## 240 **3. Experimental results and Discussions**

### 241 **3.1. Hydrate creation**

242 Figure 4 (a) shows the evolution of *FID INTENSITY* during hydrate formation (step 3)  
243 for the two tests. When methane gas is injected into the specimen, *FID INTENSITY*  
244 increases slightly during the first minutes then decreases continuously; the relationship  
245 between *FID INTENSITY* and logarithm of time during the decrease phase can be  
246 correlated with a linear function. After  $t = 40$  h *FID INTENSITY* remains constant. The  
247 results obtained by the two tests look similar even if during the first test, the data were  
248 not recorded during the first minutes. The increase of *FID INTENSITY* during the first  
249 minutes can be explained by the accumulation of methane gas inside the specimen  
250 when the gas pressure was increasing until it reached the target value (7 MPa), see  
251 Figure 4 (b) where gas pressure is plotted versus elapsed time for Test 2 (data for Test  
252 1 was not available). When the gas pressure exceeds the conditions required to create  
253 gas hydrate (3 MPa at 2 °C), gas hydrates start to be created inside the specimen.

254 This phenomenon decreases the quantity of water and increases the quantity of gas  
255 hydrates. That explains why gas hydrate formation decreases the total *FID*  
256 *INTENSITY*. Note that the intensity related to gas hydrate is negligible [31].

257

258 The following equation is used to estimate hydrate saturation ( $S_h$ ):

259

$$S_h = 1.1 \frac{I_0 + I_m - I}{I_0 + 0.1 \frac{S_{wo}}{1 - S_{wo}} I_m} S_{wo} \times 100\%$$

260 Where  $I_0$  is the initial FID intensity of the moist sand specimen and  $S_{wo}$  is the initial  
261 water saturation ( $S_{wo} = 25\%$ ). The remaining void (about 75% of total void) contents  
262 methane gas (at 7 MPa of pressure when this pressure is reached). For this reason,  $I_m$   
263 - the FID intensity of methane gas (at 7 MPa of pressure) in the specimen before the  
264 hydrate formation, equals to 75% of the value obtained from the calibration test (case  
265 (b)):  $I_m = 0.75 \times 1000 = 750$ . This equation is applicable only when the gas pressure  
266 equals 7 MPa. The underlying assumption for this equation is that water reacts locally  
267 to form hydrates, and that gas can go in and out of the sample to occupy the remaining  
268 space between hydrates and remaining water, owing to the 10% of volume increase  
269 when water is converted to hydrate. As a result, during the hydrate formation, the  
270 remaining void containing methane gas is  $(100 - S_{wo} - S_h)\%$  of total void.

271

272 Figure 4 (c) shows the estimated hydrate saturation evolution for Test 2. Hydrate starts  
273 to be created immediately when the gas pressure is higher than 3 MPa. As mentioned  
274 before, hydrate saturation is only calculated from when pore pressure reaches 7MPa  
275 (at 0.06<sup>th</sup> hour, hydrate saturation is 0.3%). The hydrate content increases then linearly  
276 with the logarithm of time and reaches its maximal value after 40 h. Note that, after 40  
277 h,  $S_h = 27\%$ , that means all water in the specimen has been transformed to hydrate,

278 and that the remaining NMR signal at the end of the process is that of the methane  
279 phase.

280

281 Figure 5 plots the signal (*i.e.* owing to water and methane) versus the elevation ( $Z = 0$   
282 corresponds approximatively to the bottom of the specimen) for various times. It can  
283 be noted that the signal is generally homogenous along the specimen elevation. At the  
284 beginning ( $t = 0$ ), the specimen contains only water and air in the pore space. Slight  
285 fluctuation of the signal along the elevation should correspond to the compaction  
286 procedure (moisture sand tamped by layers of 10 - 20 mm), which induces slight  
287 heterogeneity of porosity and water distribution in the specimen. When methane gas  
288 is injected into the specimen, GH is formed and the water content decreases  
289 progressively. That explains why the signal decreases progressively with time and the  
290 profile becomes more homogenous.

291

292 Bagherzadeh et al. [33] found that hydrate formation occurs faster in a bed with lower  
293 initial water saturation and as opposed to the higher water saturation case, hydrate  
294 formed homogeneously at 25% of initial water saturation. This is in agreement with  
295 homogenous  $Z$  profiles during GH formation in gas saturated media in this study.

296

297 Rydzy [34] investigated the kinetics of methane hydrate formation in unsaturated  
298 Ottawa sand via the combination of wave velocity measurements and MRI (Mean  
299 Intensity,  $MI$ ). The results showed that at low initial water saturation,  $MI$  decreased  
300 quickly few hours after gas injection which means hydrate saturation increased quickly.  
301 In addition, velocities (compressional and shear velocities) increased quickly during  
302 hydrate saturation, slowed down and became stable from when hydrate formation was

303 almost finished. This could be explained by cementation model illustrating hydrate  
304 growth in capillary water at sand grains contacts [37,38] which supposed that films of  
305 hydrates are first formed quickly at water-gas interfaces, the subsequent hydrate  
306 formation (from the films toward the centers of grain contacts) is slower depending on  
307 the diffusion of methane molecules through the hydrate film and water. This can be  
308 also used to explain the linear relationship between *FID INTENSITY* and logarithm of  
309 time observed in the present study during the hydrate formation. However, in the work  
310 of Rydzy [34], 5% to 12% of pore water was found to remain unconverted to hydrate  
311 at the end of experiments while in the present study almost 100% of water becomes  
312 hydrates. Actually, in the work of Rydzy [34], the signal of methane gas was not  
313 considered and that would induce errors in water content estimation in the specimen.

314

### 315 **3.2. Water Saturation**

316 Figure 6 plots *FID INTENSITY* during the water saturation (Step 4).  $t = 0$  corresponds  
317 to the opening of the valve  $V_2$ . That induces a quick decrease of *FID INTENSITY* to 0.  
318 Afterward, when water is injected to the specimen ( $t = 0.06$  h) *FID INTENSITY*  
319 increases quickly and reaches the maximal value when water pressure reaches 7 MPa.  
320 Water injection in Test 1 is slowed down between 0.1 – 0.4<sup>th</sup> hour due to an interruption  
321 to inject more water into the volume/pressure controller (12).

322

323 The signal versus elevation is plotted at various times for the Test 2 in Figure 7. Profile  
324 at 60s ( $t = 0.047$  h) was measured when methane gas was decreased to the  
325 atmospheric pressure. When water is injected from the bottom inlet, signal at the  
326 bottom starts to increase first ( $t = 0.063$  h). When the water pore pressure reaches 7  
327 MPa, the specimen can be expected to be fully saturated with water,  $t = 55$  h (methane

328 gas should not exist in this conditions). At this state, higher signal can be observed in  
329 the zone close to the bottom of the specimen while it is lower in the zone close to the  
330 top.

331  
332 Figure 8 plots the signal versus elevation for both two tests at the end of the water  
333 saturation step. The results of the Test 1 show also a higher signal close to the bottom  
334 but the signal at the top is similar to the remaining part of the specimen. The  
335 heterogeneity of water distribution at the end of this step can be explained by the  
336 saturation procedure. Actually, methane gas evacuation and water injection were  
337 performed both from the bottom. Methane gas evacuation, even if it was performed  
338 quickly, less than one minute, would induce hydrate dissociation at the zone close to  
339 the bottom. That explains why in the end, hydrate saturation at the bottom was lower  
340 (higher signal) than the other parts of the specimen.

341  
342 Hydrate saturation is estimated 20.5% and 27.5% respectively for two tests based on  
343 intensity of water saturated sample at 7MPa. Hydrates dissociation at the zone close  
344 to the bottom induces eventually the decrease of hydrate saturation compared to that  
345 after hydrate creation step. However, it is the same for Test 2. When water is injected  
346 from the bottom, the remaining gas would cumulate in the zone close to the top of the  
347 specimen, thus impeding total water saturation. At the end of water saturation phase,  
348 remaining gas would be transformed to gas hydrates, hydrate saturation in increased.  
349 That is why the hydrate saturation in this zone seems higher than in the other parts  
350 (lower signal) in Figure 8.



### 351        **3.3.        GH Dissociation-Reformation**

352    As mentioned above, a temperature cycle was performed after the water saturation  
353    phase. Figure 9 shows the pore pressure (a), the cryostat temperature (b) and *FID*  
354    *INTENSITY* (c) versus elapsed time for Test 1. It should be noted that specimen  
355    temperature could not be measured during these tests in the MRI system. However,  
356    preliminary tests performed outside the MRI system showed a characteristic time of 20  
357    min for the temperature exchange between the cryostat and the specimen. At the  
358    beginning of the tests, the pore pressure is first decreased to 4 MPa for a faster  
359    heating-induced gas hydrate dissociation as gas hydrates are closer to equilibrium  
360    boundary. Note that heating gas hydrates in undrained conditions induced the increase  
361    of pore pressure (close to corresponding equilibrium pressure of gas hydrates) mainly  
362    due to thermal dilation of water and hydrate dissociation [39]. Afterward, the cryostat  
363    temperature is increased from 2 °C to 25 °C with a constant rate. As heating is  
364    performed under undrained conditions (the valves  $V_1$  and  $V_2$  were closed), pore  
365    pressure increases according to heating and stabilizes at 14 MPa when the  
366    temperature reaches 25 °C. Heating-induced pore pressure increase is mainly due to  
367    thermal dilation of water and hydrate dissociation [39]. After this phase, the valve  $V_1$  is  
368    opened to connect the cell to the pressure/volume controller (No. 12 in Figure 2) in  
369    order to impose a pore pressure of 19 MPa. This pressure is maintained until the end  
370    of the cooling-induced hydrate re-formation phase (Step 6). At  $t = 7.5$  h, the cell  
371    temperature is decreased quickly to 2 °C to re-create GH.

372

373    Figure 9 (c) plots *FID INTENSITY* versus elapsed time during these steps. The data  
374    from the beginning to  $t = 1.6$  h was unfortunately not available. From  $t = 1.6$  h, *FID*  
375    *INTENSITY* decreases as the specimen temperature increases. Note that owing to

376 Curie law for spin polarization in the MRI magnet, *FID INTENSITY* must be here  
377 considered to be additionally influenced by temperature, being inversely proportional  
378 to its absolute value in Kelvin. For a given fluid content, it then increases when  
379 temperature decreases and vice versa. At  $t = 2.8$  h, *FID INTENSITY* starts to increase  
380 when the signal of water creation (from dissociated GH) was higher than that induced  
381 by temperature increase. In the present study, no direct temperature measurement  
382 was available inside the specimen, and no temperature correction of *FID INTENSITY*  
383 was made. At  $t = 3.6$  h, *FID INTENSITY* decreases when GH has been completely  
384 dissociated (pore pressure reached 14 MPa) but the specimen temperature continues  
385 to increase to reach the imposed temperature in cryostat. At  $t = 4.4$  h, increasing pore  
386 pressure from 14 MPa to 19 MPa induces an increase of *FID INTENSITY*. When the  
387 cryostat temperature is decreased quickly ( $t = 7.5$  h), the temperature of the specimen  
388 decreases progressively inducing an increase of *FID INTENSITY*. At  $t = 8.2$  h, GH  
389 starts to be re-created progressively inducing decrease of *FID INTENSITY*. When the  
390 GH re-creation is completed, *FID INTENSITY* stabilizes.

391  
392 The results of the Test 2 are shown in Figure 10. After reducing the pore pressure from  
393 7 MPa to 4 MPa, the cryostat temperature is increased quickly from 2 °C to 20 °C ( $t =$   
394 0.1 h) and then to 25 °C ( $t = 2.1$  h). It is decreased to 2 °C at  $t = 22$  h. Heating under  
395 undrained conditions induces an increase of pore pressure from 4 MPa to 15 MPa.  
396 The subsequent heating (from 20 °C to 25 °C) does not influence the pore pressure.  
397 From  $t = 4.1$  h, the pore pressure is maintained at 19 MPa as the case of the Test 1.  
398 The results on *FID INTENSITY* show phenomena similar to that observed in Test 1:  $t =$   
399  $0 - 0.6$  h, *FID INTENSITY* decreases due to heating;  $t = 0.6 - 1.9$  h, *FID INTENSITY*  
400 increases due to GH dissociation;  $t = 1.9 - 4$  h, *FID INTENSITY* decreases due to

401 heating; from  $t = 4$  h, *FID INTENSITY* increases due to increase of pore pressure (from  
402 14 MPa to 19 MPa);  $t = 22$  h *FID INTENSITY* increases first due to cooling then  
403 decreases due to GH re-formation. More regular *FID INTENSITY* acquisitions between  
404 2-5<sup>th</sup> hour are not available to reflect better the GH dissociation – reformation.

405

406 Figure 11 shows the signal versus elevation for the two tests at the end of the water  
407 saturation, GH dissociation, and GH reformation phases. The results show a slight  
408 redistribution of water after the GH dissociation/reformation cycle. At the end of this  
409 cycle, water seems distributed more homogeneously. Min/Mean and Max/Mean profile  
410 signal of Test 1 are (0.92; 1.30), (0.93; 1.10) respectively for water saturation and GH  
411 reformation case. Similarly, they are (0.76; 1.32), (0.90; 1.26) for Test 2.

412

### 413 **3.4. Depressurization-induced hydrate dissociation**

414 To observe the depressurization-induced GH dissociation, pore pressure is first  
415 decreased from 19 MPa to 5 MPa while specimen temperature is maintained at 2 °C.  
416 Note that, these conditions are inside the GH stabilization zone. The valve  $V_2$  is then  
417 connected to the system (9) while the valve  $V_1$  is closed. That reduces pore pressure  
418 directly to atmospheric pressure. The quantity of dissociated methane gas measured  
419 by the system (9) is used to estimate the hydrate saturation  $S_h$  remaining in the  
420 specimen. MRI data are disregarded for such purpose because ice is likely to appear  
421 in the specimen at this step and impede the direct interpretation of signal intensity.

422

423 Figure 12 shows hydrate saturation and *FID INTENSITY* versus elapsed time during  
424 the GH dissociation for Test 1 (a) and Test 2 (b). The results of Test 1 show a quick  
425 decrease of  $S_h$  from 21% at the beginning to 0 almost after 0.2 h. During this period,

426 *FID INTENSITY* decreases quickly. Once the hydrate dissociation is finished, *FID*  
427 *INTENSITY* increases slowly during the next hour. The results of Test 2 show similar  
428 trends but *FID INTENSITY* decreased more slowly at the beginning. In fact, in the  
429 objective of decelerating the gas hydrate dissociation, for Test 2, valve  $V_2$  was opened  
430 partly at the beginning (0 - 0.067<sup>h</sup> hour). However, hydrate dissociation was stopped  
431 as created gas and water were blocked in the sample. Valve  $V_2$  was so opened  
432 completely, *FID INTENSITY* decreased fast afterward. The decrease of *FID*  
433 *INTENSITY* during the hydrate dissociation phase can be explained by the expellee of  
434 water from the specimen by the created methane gas. At the same time, as hydrate  
435 dissociation is an endothermic process, ice would be formed during this phase. That  
436 induces decrease of *FID INTENSITY* even when gas hydrates are almost dissociated.  
437 In the subsequent phase, ice melting increases the quantity of liquid water in the  
438 specimen, which explains the increase of *FID INTENSITY*.

439

440 The signal versus elevation is plotted for various times during this step in Figure 13.  
441 These results confirm the statement above. Ice formation takes place only in the zone  
442 where hydrate is present (that means along the specimen except the zone close to the  
443 bottom). For this reason, signal at this zone increases at the end of the dissociation  
444 phase (which corresponds to ice melting) while the signal at the zone close to the  
445 bottom remains constant. Actually, rapid dissociation by depressurizing the sediments  
446 below the quadruple point of methane hydrate drops the temperature below the  
447 freezing point of water causing ice formation [9,10]. Heat of hydrate dissociation is 450  
448  $\text{Jg}^{-1}$  [40] while it is  $-342\text{Jg}^{-1}$  for the transformation of water at 2 °C to ice. Depending on  
449 heat transfer in the temperature control system to compensate the temperature  
450 decrease due to GH dissociation, GH reformation and ice formation ratio vary

451 depending also on the kinetics of GH dissociation. That is why, pore pressure is  
452 reduced from 19 MPa to 5 MPa before finally set up at atmospheric pressure to better  
453 observe the GH dissociation. Fan et al. 2017 [35] investigated the methane hydrate  
454 dissociation in glass beads by depressurization method. Ice formation was also  
455 observed by a rapid reduction of  $M/I$  and water distribution variation with time in the  
456 case where pore pressure was reduced below the quadruple point of methane  
457 hydrates.

458

459 To exploit natural gas hydrates after the depressurization method, the pressure in a  
460 bottom hole is first lowered by a submersible pump. During the GH dissociation, GH  
461 saturation decreases, low pressure is transferred to a distant region from well due to  
462 dramatic increase of permeability. GH dissociation stops when reservoir temperature  
463 is lower or identical to the corresponding GH equilibrium due to an endothermic  
464 reaction [14]. GH reformation and/or ice formation during GH dissociation is a common  
465 problem to overcome to increase the potential of hydrate production after the  
466 depressurization method. Some reservoir simulators (Hydrosim, MH 21, STOMP-HYD,  
467 CMG-STARs, TOUGH + HYDRATE) have been developed and are commonly used  
468 [14]. However, field scale production tests are needed to improve the accuracy of  
469 numerical predictions. In this study, due to the limited laboratory specimen size, the  
470 high production pressure and the fast depressurizing rate, the dissociation and ice  
471 formation are observed almost homogenous along the elevation. Experimental scale  
472 is then one of the important factors needed to be paid attention for future laboratory  
473 GH dissociation studies.

## 474 **Conclusion**

475 MHBS is firstly created by pressurizing methane gas (at 7 MPa) into already chilled  
476 moistened packed sand specimen (after excess gas method). Following the hydrate  
477 formation, water is injected into the specimen and the remaining gas is bled out  
478 simultaneously. A subsequent heating/cooling cycle is applied in order to completely  
479 dissociate GH and then recreate them inside the specimen. Methane hydrate  
480 dissociation after the depressurization method is also investigated after the whole GH  
481 formation process. From MRI measurements, the following conclusions can be drawn:

- 482 - Pressurizing methane gas into already chilled moistened packed sand  
483 specimen creates GH homogenously in the specimen. The formation is fast at  
484 the beginning, slows down after some hours and then stabilizes after some ten  
485 hours.
- 486 - Subsequent water saturation redistributes GH in the specimen.  $S_h$  at the water  
487 inlet is smaller than the other part (due to GH dissociation) while  $S_h$  at the  
488 opposite end could be higher (due to additional GH formation).
- 489 - Undrained heating/cooling cycle makes the GH distribution more homogenous  
490 in the specimen.
- 491 - The ice formation due to depressurization-induced GH dissociation below the  
492 quadruple point of methane hydrate is observed.

493 The findings of the present work would be helpful for future studies on MHBS in  
494 laboratory. The temperature cycle is considered as an essential step to reproduce  
495 natural MHBS homogenously in the specimen. MRI is a good mean to investigate the  
496 hydrate dissociation.

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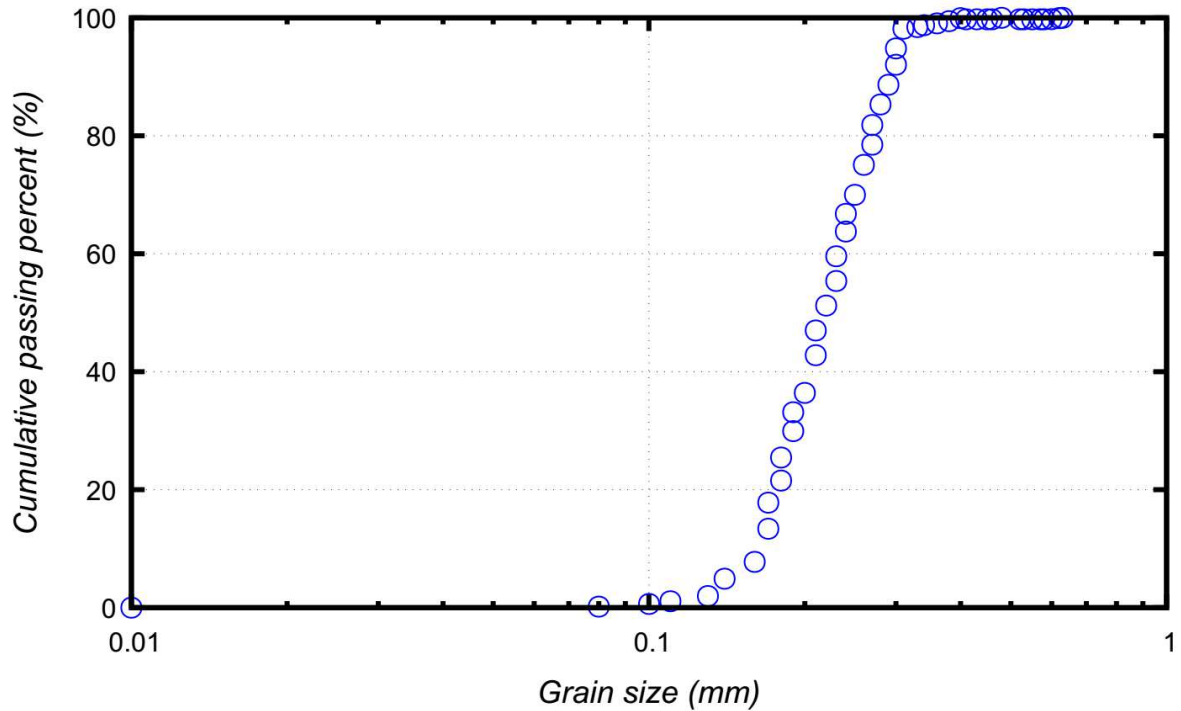
**Table 1. Characteristics of Fontainebleau sand [41]**

Material	$d_{50}$ (mm)	$d_{10}$ (mm)	$e_{min}$	$e_{max}$	Angularity	$\rho_s$ (Mg/m <sup>3</sup> )
Fontainebleau sand	0.206	0.150	0.56	0.88	Sub-rounded	2.65

633  $d_{10}, d_{50}$ : grain sizes corresponding to 10, 50 % passing respectively;  $e_{min}, e_{max}$ : minimum  
634 and maximum void ratio respectively;  $\rho_s$  grain mass density.

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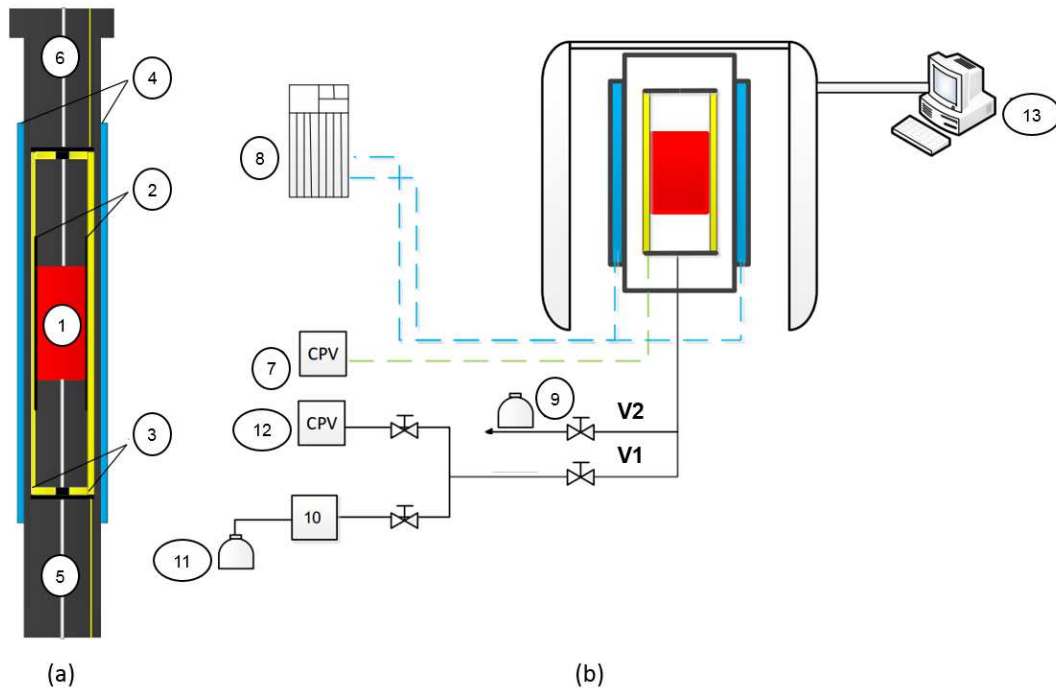


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**Figure 1. Particle size distribution (modified from [37])**

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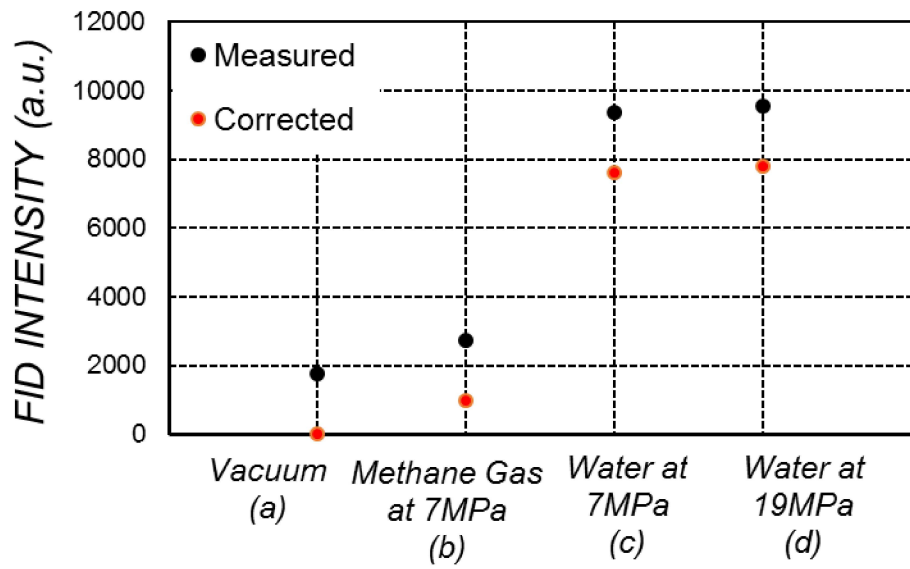
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642 1 - Sand specimen; 2 - Neoprene membrane; 3 - Confining fluid; 4 - Temperature  
 643 controlling fluid; 5 - Bottom inlet; 6 - Top inlet; 7 - Confining CPV; 8 - Cryostat; 9 -  
 644 System to measure volume of gas; 10 - Gas flowmeter; 11 - CH<sub>4</sub> bottle; 12 - Water  
 645 CPV; 13 - MRI measured system.

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**Figure 2. Schematic diagram of the experimental setup**

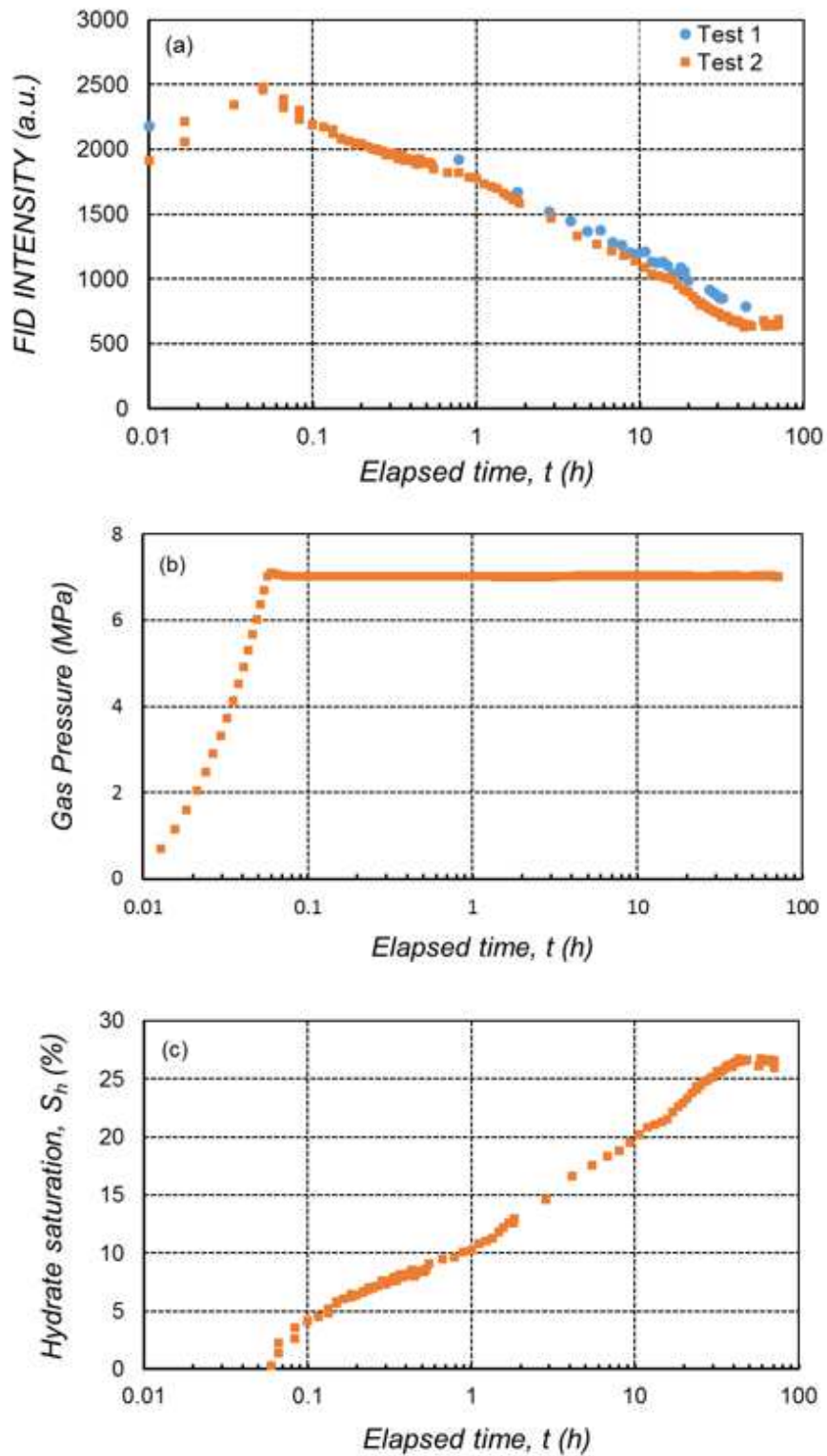
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**Figure 3. Reference signal**

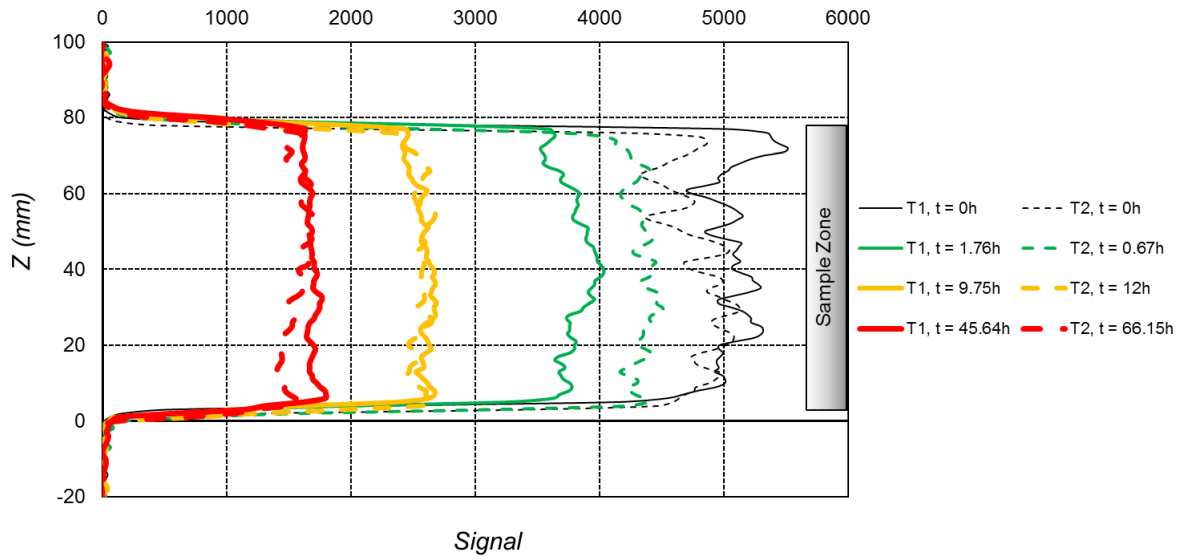


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651 **Figure 4. (a) FID Intensity evolution of the two tests during GH Formation in gas**  
 652 **saturated media; (b) Pore Pressure and (c) Estimated gas hydrate saturation**  
 653 **evolution of Test 2.**

654

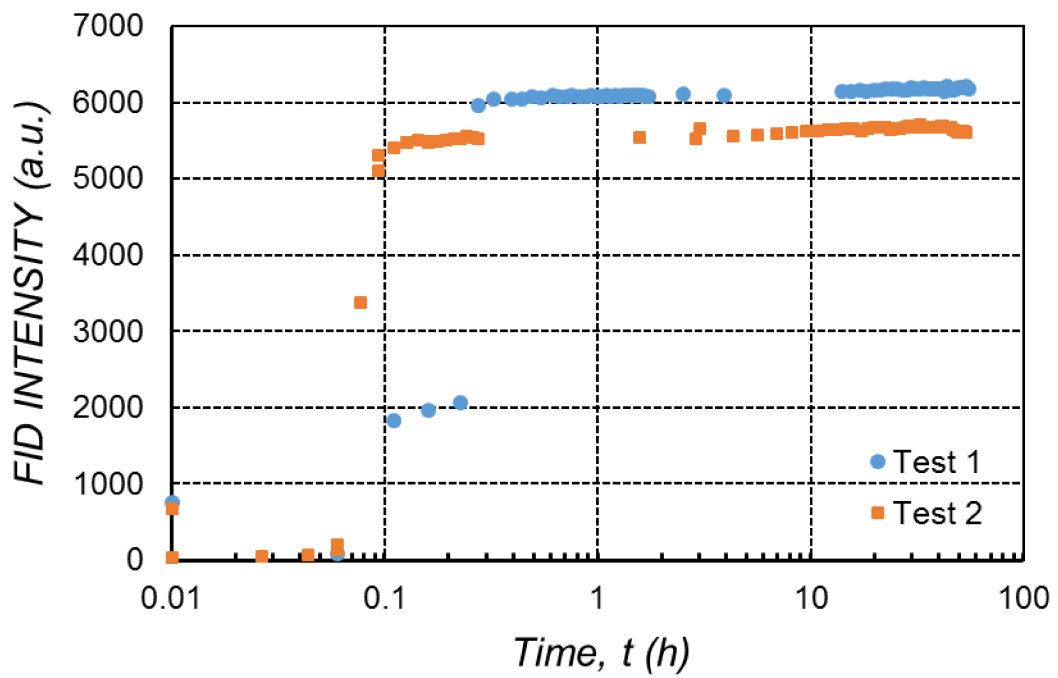
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657 **Figure 5. Signal versus elevation for the two tests during GH formation in gas**  
 658 **saturated media**

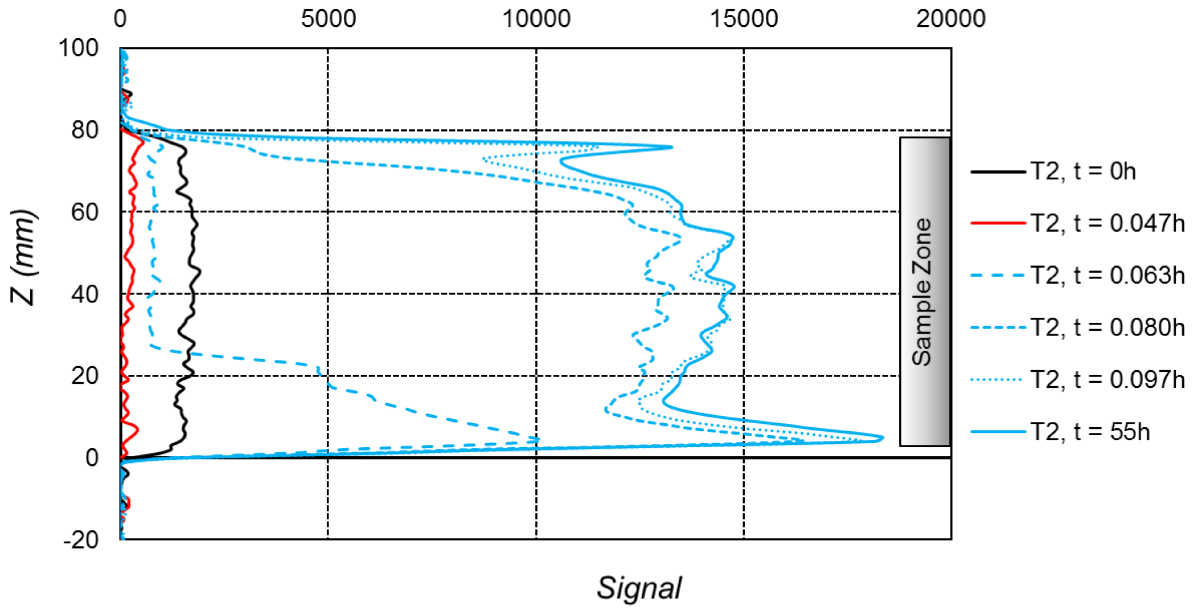
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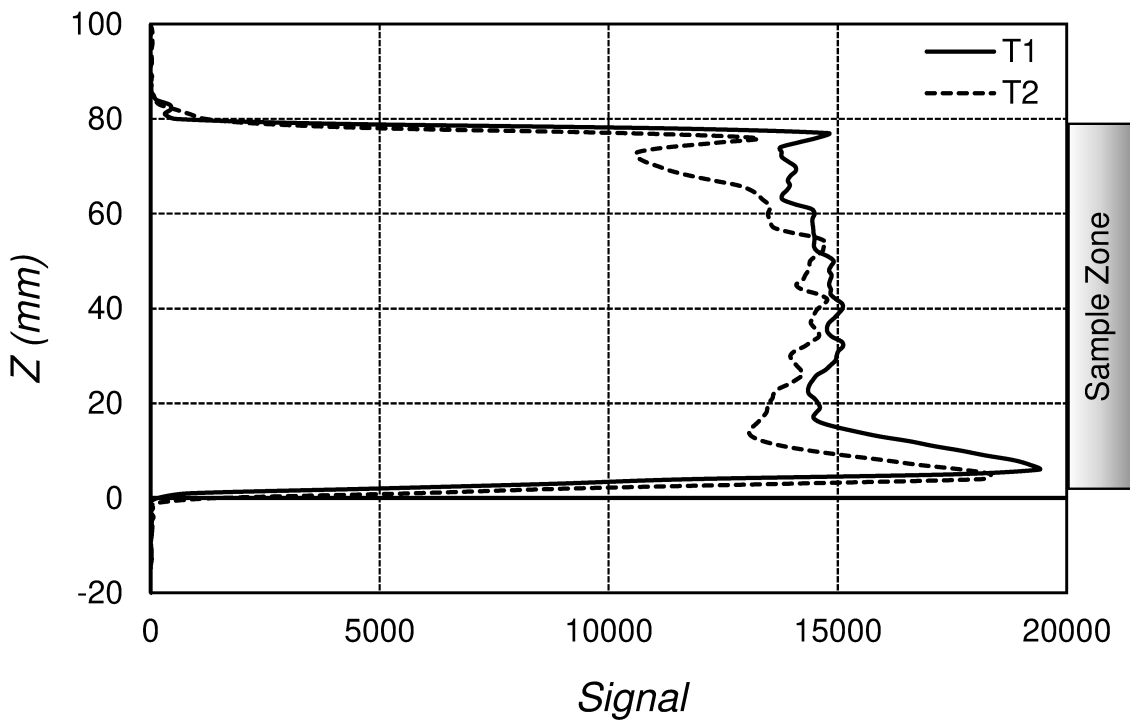
661 **Figure 6. FID Intensity evolution of the two tests during the water saturation**  
 662 **process**

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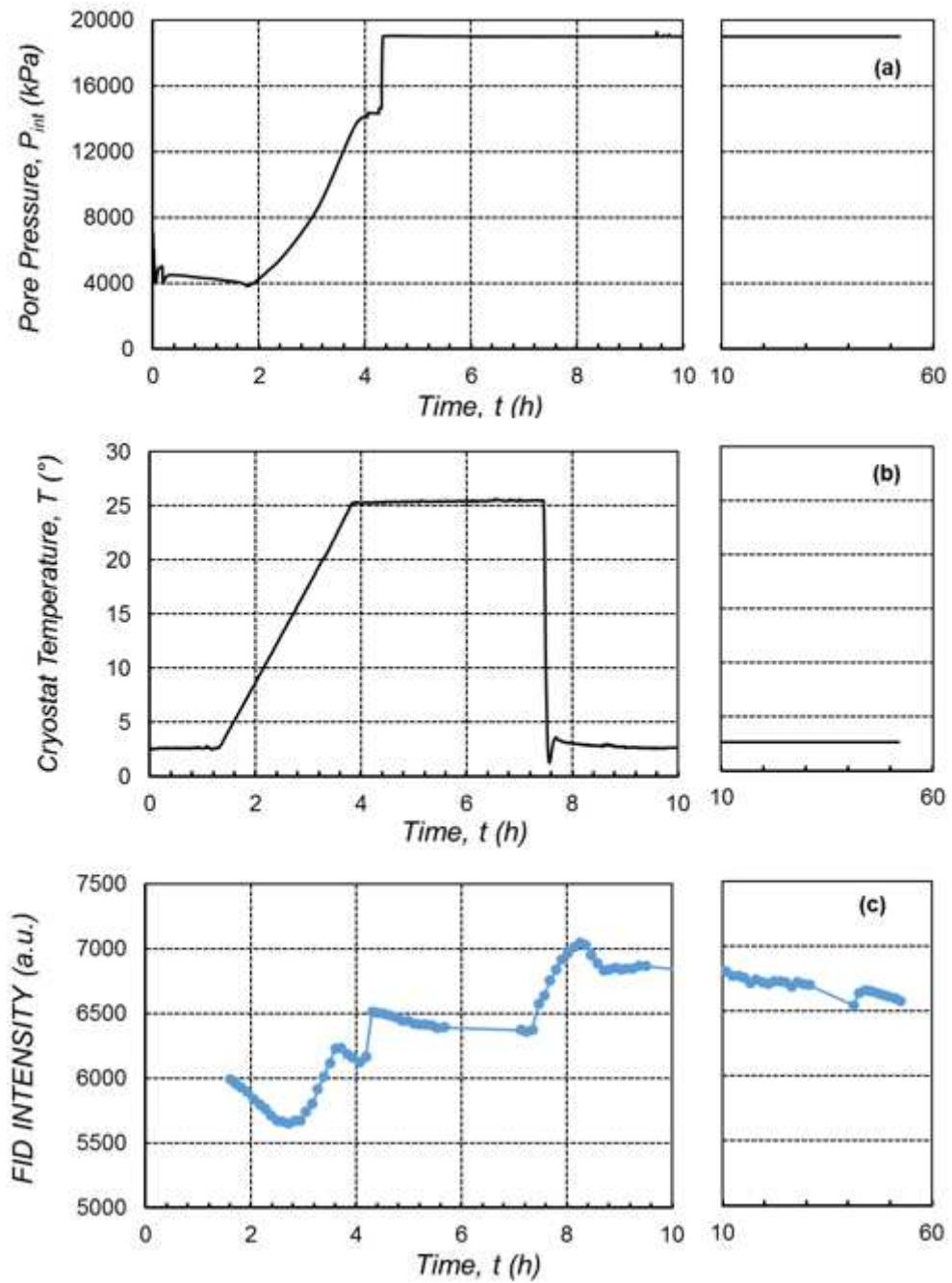
666 **Figure 7. Signal versus elevation for Test 2 during the water saturation process**



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**Figure 8. Signal versus elevation for the two tests at the end of the water saturation phase**

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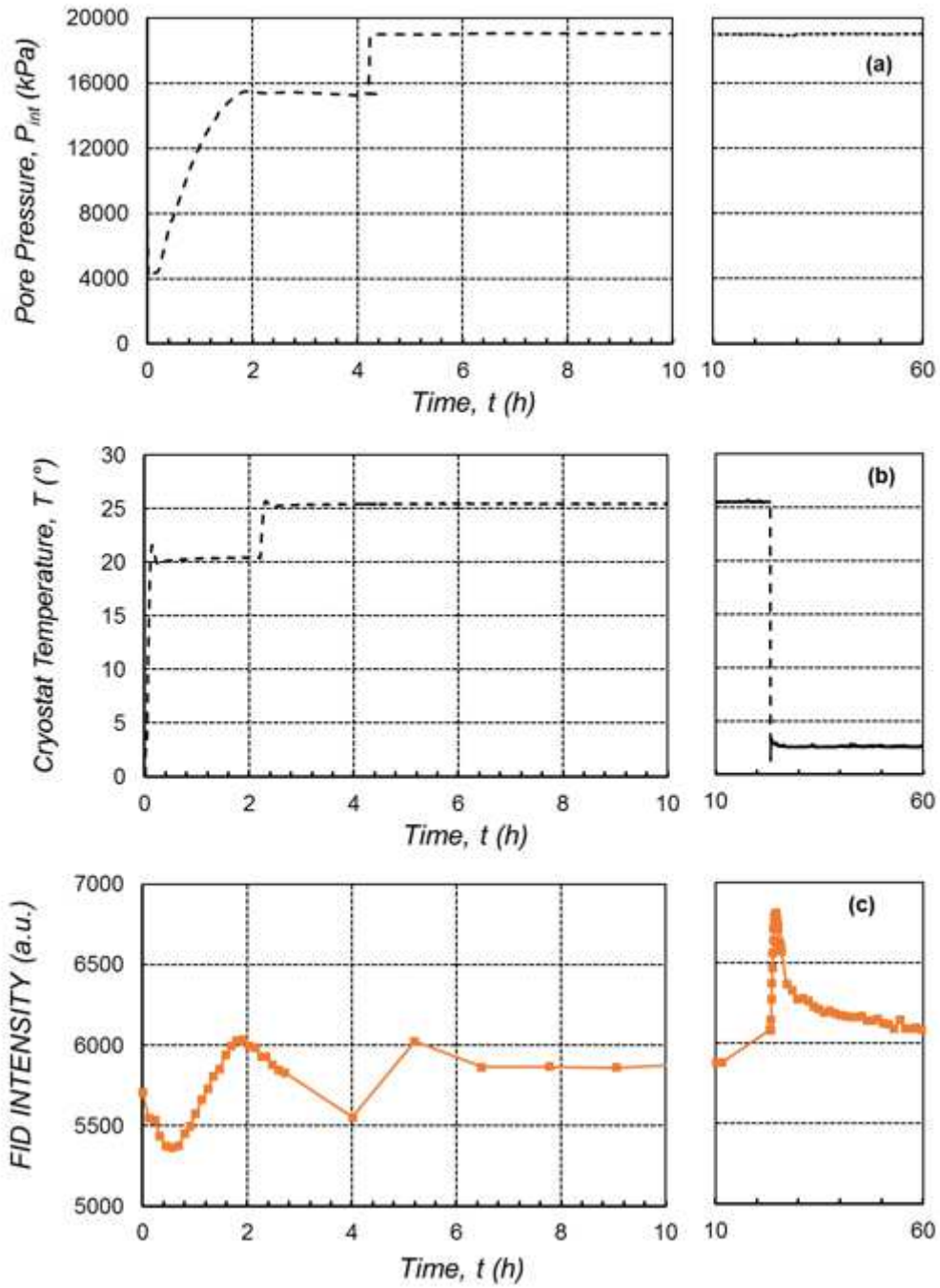
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**Figure 9. (a) Pressure evolution; (b) –Temperature evolution; (c) FID Intensity evolution during GH dissociation-reformation of Test 1**

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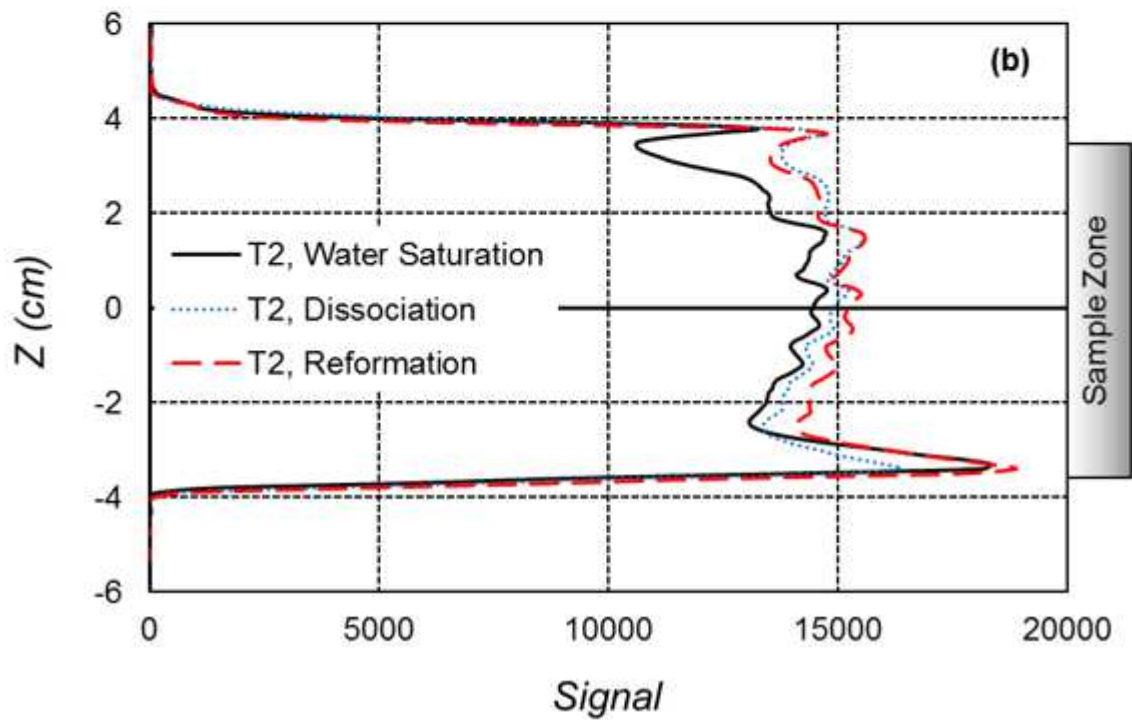
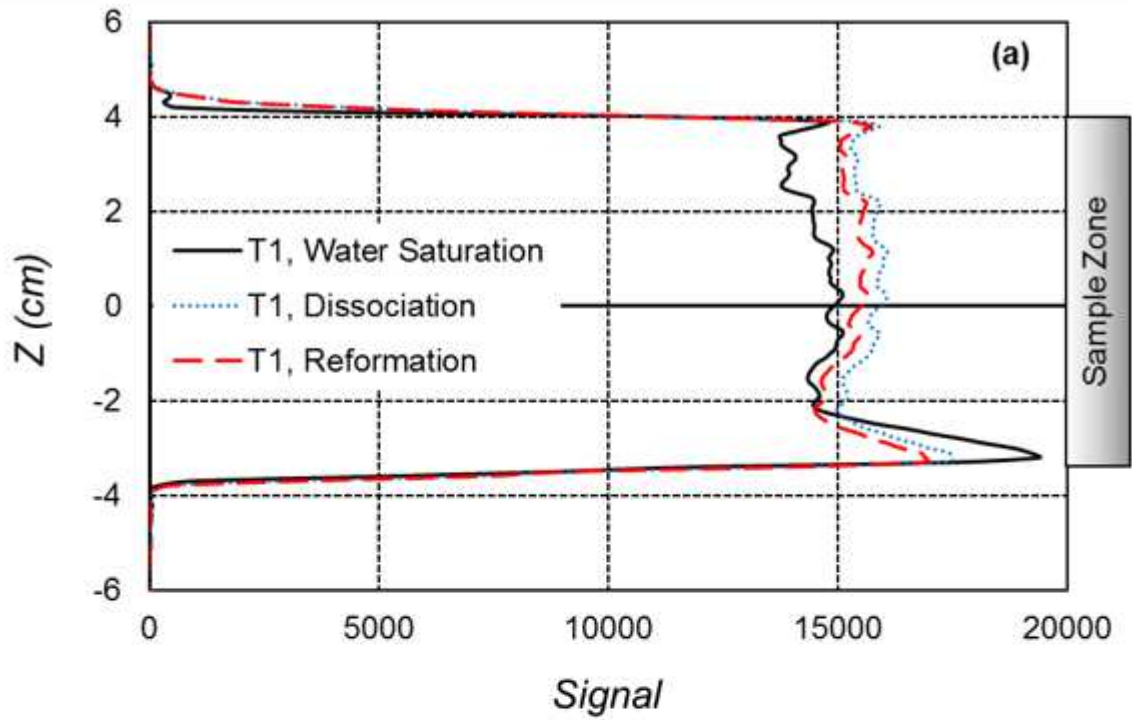
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677 **Figure 10. (a) Pressure evolution; (b) Temperature evolution; (c) FID Intensity**  
 678 **evolution during GH dissociation-reformation of Test 2**

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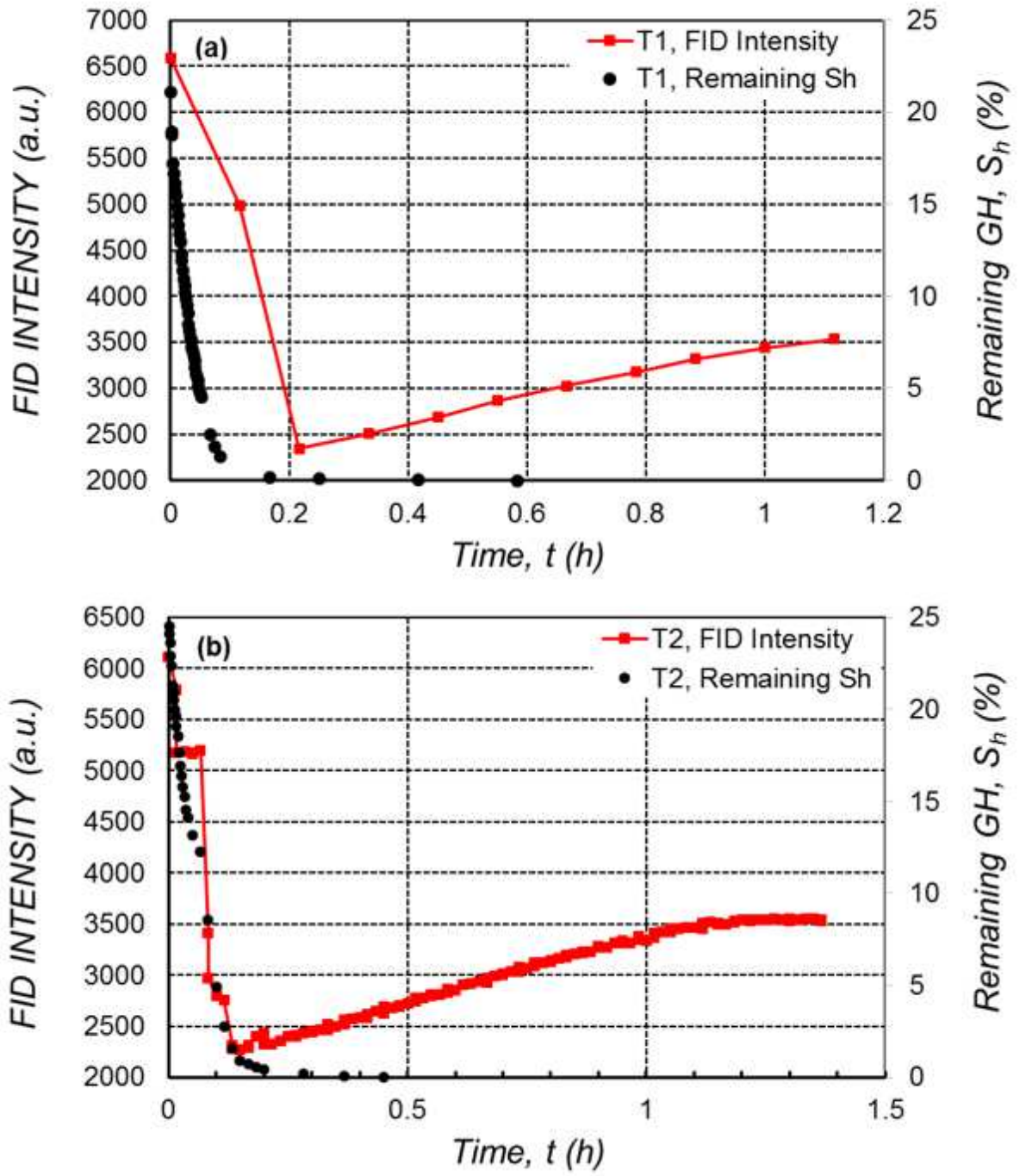
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Figure 11. Signal versus elevation at the end of the water saturation, GH dissociation, and GH reformation phases: (a) Test 1; (b) Test 2.

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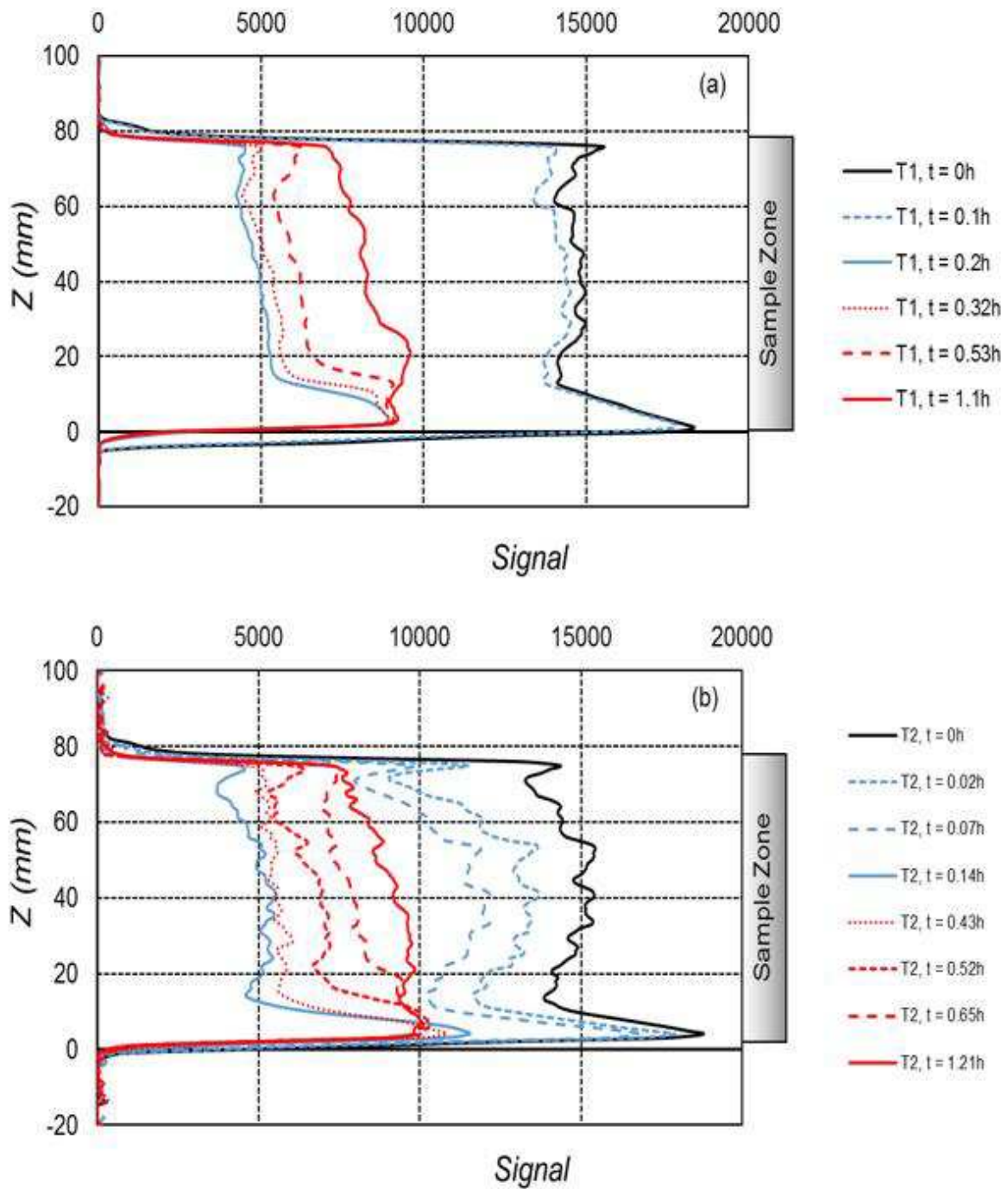
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Figure 12. FID Intensity and Remaining GH evolution during GH dissociation for (a) Test 1 and (b) Test 2.

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Figure 13. Signal versus elevation for the two tests during GH dissociation